Scholars Journal of Agriculture and Veterinary Sciences

Sch J Agric Vet Sci 2014; 1(1):28-32 ©Scholars Academic and Scientific Publishers (SAS Publishers) (An International Publisher for Academic and Scientific Resources) e-ISSN 2348-1854 p-ISSN 2348-8883

DOI: 10.36347/sjavs.2014.v01i01.005

Mass Production and Economics of Entomopathogenic Fungus, *Beauveria bassiana*, *Metarhizium anisopliae* and *Verticillium lecanii* on agricultural and industrial waste

C. S. Prasad¹, Rishi Pal²

¹ Professor O/IC, Biocontrol Laboratory(Department of Entomology) Sardar Vallabh bhai Patel University of Agriculture & Technology, Meerut-250 110 (Uttar Pradesh), India

² F.S. Biocontrol Laboratory (Department of Entomology) Sardar Vallabh bhai Patel University of Agriculture & Technology, Meerut-250 110 (Uttar Pradesh), India

*Corresponding Author

Name: Rishi Pal

Email: rishipal.biocontrol@gmail.com

Abstract: Beauveria bassiana (Balsamo) Vuellemin, Metarhizium anisopliae (Metschnikoff) Sorokin and Vertillium lecanii are entomopathogenic fungi (EPF) that occur naturally in the environment and have a potential as biological control agents against many insect pests. A laboratory study was conducted to mass produce entomopathogenic fungus, Beauveria bassiana, Metarhizium anisopliae and Verticillium lecanii on nine different agricultural and industrial waste substrates. The maximum yield (278.75 X 10⁶ spores/ml) of B. bassiana, 171.75 X 10⁶ spores/ml of M. anisopliae and 185 X 10⁶ spores/ml of V. lecanii was obtained in Farm Yard Manure (FYM) followed by SDB (246.25, 157.25 and 180.0 X 10⁶ spores/ml, respectively. The lowest number of spores of B. bassiana, M. anisopliae and V. lecanii was obtained in sugarcane bugasse (65.25, 34.25 and 39.0 X 10⁶ spores/ml), respectively. The economics of B. bassiana, M. anisopliae and V. lecanii production was evaluated based on the final yield. Among in vitro produced media, the production of spores, FYM was the best low cost substrate (Rs. 0.275, 0.440 and 0.410) for 1X 10⁶ spore production followed by Pressmud + FYM (1:1) + 1.0g dextrose (Rs. 0.39) for B. bassiana, while SDB (Rs. 0.68) for M. anisopliae and Crushed jowar grain + 1.0g dextrose (Rs. 0.52) for V. lecanii. The highest cost of spore production was recorded in sugarcane bagasse (Rs. 1.14, 2.18 and 1.92) followed by pressmud for B. bassiana, M. anisopliae V. lecanii.

Keywords: Beauveria bassiana, Metarhizium anisopliae, Verticillium lecanii, mass production, economics.

INTRODUCTION

The global consensus to reduce inputs of chemical pesticide which are perceived as being hazardous by some consumers has provided opportunities for the development of novel, benign, sustainable crop protection strategies. A great many chemical pesticides have been or are being phased out (e.g. organochlorine insecticides, methyl bromide) either because of potential human health risks, environmental pollution, effects on non-target organisms or the development of pest resistance.

Biological control agents such as entomopathogenic fungi (EPF) can be used as a component of integrated pest management (IPM) of many insect pests. Under natural conditions, these pathogens are a frequent and often cause natural mortalities of insect populations. Several fungal species including *Metarhizium anisopliae*, *Verticillium lecani* and *Beauveria bassiana* are being used as biocontrol agent for a number of crop, live stock and human nuisance pests [1].

Moreover, entomopathogens are cheap to mass produce, easy to store and effective over a wide range of temperatures and humidity levels. It also provides a rapid kill at economical doses and recently, the fungus has commercially been widely developed as a microbial agent for pest management [2] and encouraging results were obtained against whiteflies, aphids, thrips and mealybugs in greenhouses and nurseries [3]. B. bassiana, M. anisopliae and V. lecani are a facultative pathogen and can be mass-produced on various substrates. The choice of a suitable and economic medium, which supports rapid growth without loss of virulence for number of generations, is one of the basic requirements in the mass production of fungi for microbial control of insect pests. The production can be achieved using different methodologies, which can be classified in to low input and industrial technologies. However, most production of fungal spores worldwide is carried out using simple technologies that demand low inputs[4]. Mass production of entomopathogenic fungi and testing of germination are important steps in successful utilization of EPFs as biocontrol agents.

Some attempts to mass produce *B. bassiana*, *M. anisopliae* and *V. lecani* on low cost materials have been carried out by Mazumdar *et al.* [5] and Babu *et al.* [6]. They evaluated solid and liquid substrates,

including sugar industry by-products and observed that addition of dextrose was indispensable for accelerated growth on some of them. The type of growing medium, affects conidial production of entomopathogenic fungi [7]. Hence this study was design to mass culture the entomopahogenous fungi on locally available agricultural and industrial waste for a cheap and suitable substrates for the low cost production of entomopathogens of virulent spores i.e. liquid and solid media. Despite the many attempts to screen commercially available, low-cost ingredients of industrialized biological pesticides, the research on nutritional requirements of fungal agents were overlooked and a systematic investigation of fungal nutrition utilization is much needed to improved mass production and accelerates commercialization[8]. The outcome of this research will help in mass production technology of entomopathogens in a suitable and economic medium for its dissemination in field.

MATERIALS AND METHODS

The present study was carried out in Bio-control Laboratory, S.V.P. University of Agriculture & Technology, Meerut (India) in 2010. The experiment was conducted in a laboratory to test the effect of different agricultural and industrial waste, solid and liquid substrates on mass production and economics of entomopathogenic fungi (EPFs).

Nucleus culture

The nucleus culture of *B. bassiana*, *M. anisopliae* and *V. lecani* was isolated from the infected lepidopteran larva and soil-isolated fungi from different location and orchards of Meerut district. It was thereafter maintained on Sabouraud Dextrose Agar (SDA) medium as per the procedure of Prasad *et al.* [9]. Briefly, SDA was prepared and sterilized at 121° C (15 lbs) for 20 minutes. Then cooled and poured in presterilized Petri plates and *B. bassiana* from stock culture was inoculated aseptically. The Petri plates were incubated at $25 \pm 1^{\circ}$ C in BOD incubator for two weeks to harvest the inoculums culture.

Substrates and in vitro production

Nine mass production substrates were evaluated for the conidial production of *B. bassiana*, *M. anisopliae* and *V. lecanii* under controlled conditions of $25 \pm 1^{\circ}\text{C}$ in BOD incubators for 3 weeks. The substrates were (i) FYM, (ii) Sabouraud dextrose broth (SDB), (iii) Sugar industry Press mud, (iv) Sugarcane bagasse, (v) FYM liquid + 1.0g Dextrose, (vii) Pressmud liquid + 1.0g Dextrose, (viii) FYM liquid + Pressmud liquid (1:1) + 1.0g Dextrose , (viii) *Corcyra* rearing waste (Maize) and (ix) Jawar grain + 1.0 g Dextrose. The last six medium were supplemented with dextrose. There were nine treatments in three replications.

A quantity of 130 g dehydrated SDB was suspended in 2000 ml distilled water, heated to dissolve the medium (pH 5.6) and sterilized by autoclaving at 15

lbs pressure (121°C) for 15 minutes. The lukewarm liquid media was poured in conical flask. Then *B. bassiana, M. anisopliae* and *V. lecanii* were inoculated aseptically and then conical flasks were incubated in BOD.

The Corcyra rearing waste (crushed maize grain) and sorghum grain were soaked over night, cleaned with fresh water and 100 g of each was put in separate Conical flasks (250-ml capacity) supplemented with dextrose (1.0 g), plugged with non-absorbent cotton and autoclaved. Upon cooling of media, B. bassiana, M. anisopliae and V. lecanii were inoculated aseptically and incubated in BOD incubator. The crushed jowar grain was collected from the Bio-control Laboratory as a waste material from Corcyra Rearing Unit. The crushed jowar grain (weight 50 g) was put in flask and 15 ml distilled water with 1 g dextrose was added. The sugarcane bagasse and Pressmud was procured from Modipuram market, Meerut. The sugarcane bagasse was cut into small pieces and half-filled in flask with 15 ml distilled water.

Spore counting

A drop of conidial suspension of *B. bassiana*, *M. anisopliae* and *V. lecanii* (obtained from the growing media by filtering through muslin cloth) was placed on the hemocytometer. The cover glass was put over the grid carefully so that no air bubble entered between cover glass and slide. The conidia of entomopathogenic fungi were counted under Olympus BX41 phase contrast microscope at higher resolution in the middle square (V) of hemocytometer which contained 25 groups of 16 small squares, each group 0.2 mm square.

Statistical analysis

The conidial production of entomopathogernic fungi from different substrates were subjected to analysis of variance (ANOVA) using SPSS 10.0 for Windows software (SPSS, 1999) [10]. The means were separated using LSD and differences between treatments were considered significant at p < 0.05.

RESULTS AND DISCUSSION

The optimum nutrient medium is necessary for the adequate growth of microorganism in general and fungi in particular (Altomare et al., 1999). In the present study, one chemical media (and SDB), and six waste materials (i.e., FYM, Pressmud, sugarcane bagasse, broken maize grain (*Corcyra* rearing waste) and broken jowar were evaluated *in vitro*.

Mass production of B. bassiana

The production was more on media containing FYM, SDB and FYM liquid + Pressmud liquid (1:1) + 1g Dextrose as compared to the rest of the media. The least conidial growth was observed in crushed jowar grain and sugarcane bagasse (Table 1).

The results showed that among the media tested, FYM (278.75 X 10^6 spores/ml), SDB (246.25 X 10^6 spores/ml) and FYM liquid + Pressmud liquid (1:1) + 1g Dextrose (200.25 X 10^6 spores/ml) were the most suitable media with higher growth of mycelium and production of large number of spores of *B. bassiana*. While the sugarcane bugasse (65.25 X 10^6 spores/ml) produces least number of spores followed by FYM liquid + 1g Dextrose (97.50 X 10^6 spores/ml). Pandey and Kanaujia [11] obtained 6.4×10^7 conidia/ml in

SDA medium followed by finger millet (5.4×10^7 conidia/ml) whereas Purwar and Sachan [12] recorded SDA with yeast extract and sorghum-based media producing higher biomass and conidial counts. Many earlier researcher successfully used SDA medium for the mass culture of fungi. However, surprisingly in the present study the spore count was less in SDB medium as compared to FYM. Hallsworth and Magan [13] offered SDA medium for the successful culture of *B. bassiana* and *M. anisopliae*.

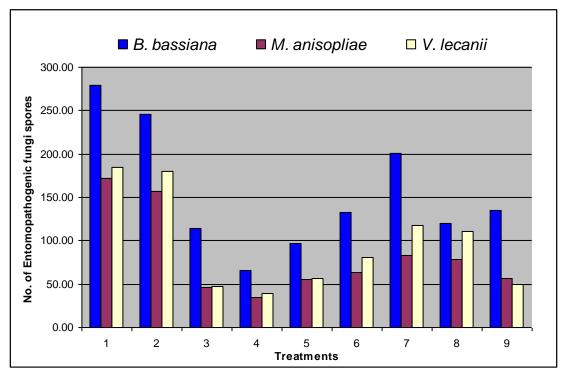


Fig. 1. Mass production of Entomopahogenic fungi on agricultural and industrial waste

Table 1. Mass production of B. bassiana, M. anisopliae and V. lecanii on different substrates.

S.	Media	Spore count (X 10 ⁶)		
No.		B. bassiana	M. anisopliae	V. lecanii
1	Farm Yard Manure (FYM)	278.75	171.75	185.00
2	Sabouraud dextrose broth (SDB)	246.25	157.25	180.00
3	Pressmud (Sugar mill)	114.00	45.75	47.50
4	Sugarcane bagasse	65.25	34.25	39.00
5	FYM liquid + 1.0g Dextrose	97.50	55.50	56.50
6	Pressmud liquid + 1.0g Dextrose	132.50	63.50	80.50
7	FYM liquid + Pressmud liquid (1:1) +1.0g dextrose	200.25	83.25	117.50
8	Corcyra rearing waste (Maize)	119.75	78.75	110.75
9	Jowar grain + 1.0 g Dextrose	135.25	57.00	49.50
CD at 5%		35.850	18.832	31.585
SE (m)		11.977	5.289	10.549

Figures are parenthesis in no transformation

^{*}Means (of three replicates where each replicate comprised of 15 individual flasks containing each substrate) \pm standard error indicated with the same letter are not significantly different according to LSD test at p < 0.05.

Table 2. Economics of mass production of *B. bassiana*, *M. anisopliae* and *V. lecanii* $(1 \times 10^6 \text{ spores})$ on different substrates of agricultural and industrial waste.

S. No.	Media	Cost of production of 1 X 10 ⁶ spores (Rs.)			
		B. bassiana	M. anisopliae	V. lecanii	
1	Farm Yard Manure (FYM)	0.27	0.44	0.41	
2	Sabouraud dextrose broth (SDB)	0.43	0.68	0.59	
3	Pressmud (Sugar mill)	0.66	1.66	1.60	
4	Sugarcane bagasse	1.14	2.18	1.92	
5	FYM liquid + 1.0g Dextrose	0.78	1.37	1.33	
6	Pressmud liquid + 1.0g Dextrose	0.57	1.20	0.94	
7	FYM liquid + Pressmud liquid (1:1) +1.0g Dextrose	0.39	0.94	0.66	
8	Corcyra rearing waste (Maize)	0.62	0.96	0.68	
9	Jowar grain + 1.0g Dextrose	0.56	1.34	1.55	
CD at 5%		0.054	0.035	0.035	
SE (m)		0.021	0.012	0.012	

Figures are parenthesis in no transformation value

Mass production of M. anisopliae

The results showed that among the media tested, FYM (171.75 X 106spores/ml), SDB (157.25 X 106 spores/ml) and FYM liquid + Pressmud liquid (1:1) + 1g Dextrose (83.25 X 10⁶ spores/ml) were the most suitable media with higher growth of mycelium and production of large number of spores of M. anisopliae. While the sugarcane bugasse (34.25 X 10⁶ spores/ml) produces least number of spores followed by Pressmud (45.75 X 10⁶ spores/ml). Pandey and Kanaujia [11] obtained 6.4×10^7 condia/ml in SDA medium followed by finger millet (5.4 \times 10⁷ conidia/ml) whereas Purwar and Sachan [12] recorded SDA with yeast extract and sorghum-based media producing higher biomass and conidial counts of B. bassiana. No any fungal spore in sugarcane bagasse + 1.0% yeast extract and pressmud + 1% yeast extract while 1.70 X 10⁴ spores was obtained in pressmud with 10% molasses reported by Bharati et al[14]. However, surprisingly in the present study the spore count was less in SDB medium as compared to FYM. Hallsworth and Magan [13] offered SDA medium for the successful culture of B. bassiana and M. anisopliae. Soundrapandian and Chandra [15] were also found 300X10⁴ spores of *M. anisopliae* in SDA.

Mass production of V. lecanii

The results showed that among the media tested, FYM (185.00 X 10^6 spores/ml), SDB (180.00 X 10^6 spores/ml) and FYM liquid + Pressmud liquid (1:1) + 1g Dextrose (117.50 X 10^6 spores/ml) were the most suitable media with higher growth of mycelium and production of large number of spores of *V. lecanii*. While the sugarcane bugasse (39.00 X 10^6 spores/ml) produces least number of spores followed by Pressmud (47.50 X 10^6 spores/ml). The reports of sorghum grains were found to be ideal for mass production of *V. lecanii* was documented by Sahayraj and Namasivayam [16]. Pandey and Kanaujia [11] obtained 6.4×10^7 conidia/ml in SDA medium followed by finger millet (5.4 \times 10^7 conidia/ml) whereas Purwar and Sachan

[12] recorded SDA with yeast extract and sorghumbased media producing higher biomass and conidial counts of *B. bassiana*. The above findings are closely agreed with Bharati *et al.*[14] who recorded no fungal spores of *M. anisopliae* in sugarcane bagasse + 1.0% yeast extract and pressmud + 1% yeast extract and also found spore yields were considerably low in agrowastes as compared to food grains.

Economics of mass production of B. bassiana, M. anisopliae and V. lecanii

Based on cost incurred for the production of spores FYM was the best low cost substrate (Rs 0.275, 0.440 and 0.410.) for 1X 10⁶ spore production followed by Pressmud + FYM (1:1) + 1.0g Dextrose (Rs. 0.39) for *B. bassiana*, while SDB (Rs 0.68.) for *M. anisopliae* and Crushed jowar grain + 1.0g dextrose (Rs 0.52.) for *V. lecanii* (Table 2). The highest cost of spore production was recorded in sugarcane bagasse (Rs. 1.14, 2.18 and 1.92) for *B. bassiana*, *M. anisopliae V. lecanii* followed by FYM liquid + 1.0g Dextrose (Rs 0.78.) for *B. bassiana* and pressmud (Rs. 1.66 and 1.60) for *M. anisopliae* and *V. lecanii*.

The methodology evaluated in this study allows for the production of high quality *B. bassiana*, *M. anisopliae* and *V. lecanii* fungal spores, but the quantity produced are only suitable for small-scale laboratory and field trails.

Acknowledgements

The authors express to financial support from Indian Council of Agricultural Research, New Delhi for the project-Niche Area of Excellence is gratefully acknowledged.

REFERENCES

 Tanda Y, Kaya HK; Insect Pathology. Academic, San Diego, CA. 1993.

- Faria M, Wraight SP; Biological control of *Bemisia tabaci* with fungi. Crop Protec., 2001;20: 767-778.
- Faria MR, Wraight SP; Mycoinsecticides and mycoacaricides: a comprehensive list with worldwide coverage and international classification of formulation types. Biol. Control, 2007;43: 237-256.
- Alves SB, Pereira RM; Production of Metarhizium anisopliae (Metsch.) Sorok. and Beauveria bassiana (Bals.) Vuill. in plastic trays. Ecossistema, 1989;14: 188-192.
- Mazumdar D, Hallsworth, Magan, Puzari KC, Hazarika LK; Mass production of *Beauveria* bassiana and its potentiality on rice hispa. Indian Phytopathol. 1995; 48: 275-278.
- Babu JCS, Venkatachalapathy CM, Anitha CN; Evaluation of locally available substrate for mass production of entomopathogenic fungi, *Metarhizium anisopliae* (Metch.) Sorokin. Journal of Biopesticides, 2008;1(2): 146-147.
- 7. El Damir M; Effect of growing media and water volume on conidial production of *Beauveria bassiana* and *Metarhizium anisopliae*. J. of Biol. Sci, 2006;6(2): 269-274
- 8. Sun MH, Liu XZ; Carbon requirements of some nematophagous, entomopathogenic and mycoparasitic Hyphomycetes as fungal bio-control agents. Mycopathologia, 2006; 162(2): 295-305.
- Prasad CS, Prasad M, Hussain MA, Pal R, Kumar P, Kumar L, Singh R; Mukhye Jaivik Pertinidhiyon

- *ka Vyasaik Utpadan Tekneek* (in Hindi: Techniques for commercial production of important bioagents). Biocontrol Lab., S.V.P.U.A.&T., Meerut, India, 2008; 72.
- 10. SPSS. Statistical product and service solution, system user's guide Version 10.0. 1999.
- 11. Pandey AK, Kanaujia KR; Effect of different grain media on sporulation, germination and virulence of *Beauveria bassiana* (Balsamo) Vuillemin against *Spodoptera litura* Fabricius larvae. J. Biol. Control, 2005;19: 129-133.
- 12. Purwar JP, Sachan GC; Evaluation of media for growth and sporulation of entomogenous fungi. Ann. Pl. Protec. Sci., 2006;14: 484-485.
- 13. Hallsworth JE, Magan N; Culture age, temperature and pH affect the polyol and trehalose contents of fungal propagules. Appl. Environ. Microbiol. 1996; 62:2435-2442.
- 14. Bharati T, Kulkarni JH, Krishnaraj PU, Alagawadi AR; Evaluation of Food Grains and Agro Wastes for Sporulation of *Metarhizium anisopliae* (Ma2). Karnataka J. Agric. Sci., 2007; 20(2): 424-425
- Soundrapandian P, Chandra R; Mass production of entomopathogenic fungi, *Metarhizium anisopliae* (Metch.) Sorokin. Research Journal of Microbiology, 2007; 2(9): 690-695.
- 16. Sahayaraj S, Mamasivayam SKR;. Mass production of entomopathogenic fungi using agricultural products and by products. African Journal of Biotechnology, 2008; 7(12): 1907: 1910.