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Growth Response and Haematological Constituents of Broiler Birds Fed Raw Sand Box (Hura Crepitans) Based Diets

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Abstract Original Research Article

Growth response and haematological indices of broiler birds of 28 week of age fed raw Hura crepitans seed meal (HSM) as partial replacement for maize and soybean was examined. Three hundred (300) broiler birds were assigned to five dietary treatments (T1, T2, T3, T4 and T5) in a completely randomized designed of 60 birds per treatment, replicated thrice with each replicate having 20 birds. The control diet had 0% Hura crepitans meal while T2, T3, T4 and T5 had 5%, 10%, 15% and 20% HSM. The raw seed meal was moderate in protein (24.80%) and crude fibre (11.82%) but very high in ether extract (39.45%). The seed was also moderate in trypsin inhibitor (0.92mg/g), Tannin (0.51mg/g), Cyanide (0.23mg/g) and saponin (0.33mg/g). The HSM produced statistically significant (p<0.05) depression in feed intake, final live weight, average weight gain, feed conversion ratio and protein efficacy from inclusion level above 10% HSM, and tended to be depressed (P<0.05) more as the inclusion levels of Hura crepitans meal increased in the diets with the least values recorded for birds on treatment diet 5(20%) Hura crepitans meal. The abdominal fat deposit inceased significantly (P<0.05) as the level of Hura crepitans meal increases in the diets. The nutrient retension showed significant (P<0.05) effect only on fibre, ether extract and protein retention among other nutrient retention parameters. Both the fibre and protein retention were significantly depressed only at 15% and 20% TSM. The organs showed no significant differences except for the liver that shrinks at the 15% and 20% inclusion levels of HSM. The haematological indices also showed significant differences, with notable reduction in Red Blood Cells (RBC), packed cell volume (PCV) and hemoglobin (Hb) but only at the 20% HSM inclusion level. It was concluded that not more than 10% raw Hura crepitans could replace parts of maize and soybean meal in broiler diets without any adverse effect on growth performance and only at 20% will the haematological parameters of broiler birds be affected.

Keywords: Hura crepitans, growth response, haematological indices.

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INTRODUCTION

In developing countries, the conventional energy and protein feedstuffs are not only scarce but are expensive. The utilization of unconventional feed resources that are less in demand by man and his industries, in a way has reduced feed cost [1, 2].

This ultimately lowers cost of livestock since cost of feed accounts for 70-80% of the total cost of poultry production [3]. Therefore has led to the search for alternative feedstuffs, that as a matter of necessity must be easily available, easy to procure, easy to be produced and processed and must have comparable cost advantage over the conventional feedstuff [4].

A species that can be exploited in this direction is Hura crepitans (sand box). It possesses as much

and nutritional potential the conventionally used energy and protein sources. The seed dry matter is between 91-95% an indication of its good long storage life which will make it devoid of moldy growth. The seed is moderate in crude protein (23-28%) but high in amino acids levels especially lysine, methionine, cystein, threomine and histidine [5]. The seed is said to be high in oil (38.95-51.24%), making it a good energy source to livestock feed. However, the seed is noted to contain some antinutritional factors like trypsin-inhibitor, tannin when consumed caused vomiting, diarrhea and cramps [6-8].

This research therefore evaluate the growth response and haematological constituents of broiler birds to diets containing different levels of raw Hura crepitans (sand box) seed meal as a partial replacement for maize and soybeans.

MATERIALS AND METHODS

Source and Processing of Hura crepitans seed

The matured sand box seeds were plucked from trees within Calabar Municipality where the experiment was also conducted. The seeds were separated from its casing (dehulled) and sundried for 6 days and finally oven dried at 120°C for 6 hours. The

seeds were further milled with a meadow model 35 hammer mills and sieve through a mesh of 5mm after passing it through ethanol as a defeating agent. The proximate composition of the test ingredients and diets were determined by the method of AOAC [9] while phytate and tannin by method of Mega [10], Alkaloids, saponin and haemagglutinins by method of Liener [11].

Table-1: Proximate composition and anti-nutritional factors of raw Hura crepitans meal

Proximate Components (%)	
Dry matter	92.40
Crude Protein	24.80
Crude fibre	9.48
Ether extract	28.04
Ash	4.81
Nitrogen Free Extract	32.87
Anti-nutritional factor (mg/g)	
Trypsin Inhibitor	0.92
Tannin	0.51
Cyanide	0.23
Saponin	0.33
Haemagglutinin(Hu/g)	188.05

Diets and Management of Birds

Five experimental diets were formulated in such a way that Diets T_1 contained 0% HSM which served as the control diet. The proportion of maize and soybean were partially replaced by 5%, 10%, 15% and 20% HSM to from Diets T_2 , T_3 , T_4 and T_5 respectively. The diets were made to be iso-caloric and isonitrogenous. Three hundred (300) day old broiler chicks of 'Hacco' strain of mixed sex were raised on a common diet (vital feed) for 28 days with all needed vaccination and medication giving before being

randomly divided equally into 5 groups of 60 birds each. Each groups had 3 replicates of 20 birds each. Feed and water were given *ad libitum* for the period of 28 days of which the experiment lasted.

At the end of the experiment 3 birds per replicate were randomly selected and separated into metabolic cages for nutrient retention experiment, after 4 days of acclimatization, feacal droppings collected and feed fed were taken for analysis.

Table-2: Gross composition of experimental diets

	Levels of inclusion of Hura crepitans meal (HCM)								
Ingredients (%)	0%	5%	10%	15%	20%				
Maize	56.50	51.50	46.50	41.50	36.50				
HCM	0.00	5.00	10.00	15.00	20.00				
SBM	27.00	29.00	31.00	33.00	35.00				
Wheat Offal	10.00	8.00	6.00	4.00	2.00				
Fish Meal	2.25	2.25	2,25	2.25	2.25				
Bone meal	3.50	3.50	3.50	3.50	3.50				
*Vit/Min Premix	0.25	0.25	0.25	0.25	0.25				
Lysine	0.25	0.25	0.25	0.25	0.25				
Methionine	0.25	0.25	0.25	0.25	0.25				
Total	100.00	100.00	100.00	100.00	100.00				
Calculated Analysis %									
Crude protein	22.01	21.97	21.94	21.90	21.88				
Crude fibre	4.50	4.52	4.58	4.61	4.69				
Available phosphorus	0.81	0.79	0.78	0.77	0.75				
Calcium	1.21	1.20	1.18	1.18	1.16				
ME(Kcal/kg)	3005.18	3011.10	3019.21	3024.50	3035.50				
Determined Analysis %									
Crude protein	21.58	21.47	21.34	21.22	21.11				
Crude fibre	4.16	4.29	4.35	4.52	4.70				
Gross energy(mg/kg)	4.09	4.11	4.13	4.16	3.19				
Ether extract	3.58	3.62	3.66	3.71	3.90				
Ash	7.81	7.62	7.50	7.41	7.37				

*Vitamin/mineral premix containing the following per kg. Vitamin A, 8,000,000 IU; vitamin D3, 1,600000 IU; Vitamin E, 5,000 IU; Vitamin K, 2,000 mg; Thiamine, 1,500 mg; Riboflavin B2, 4,000 mg; Pyridoxine B6, 1,500 mg; Antioxidant, 125 g; Niacin, 1,500 mg; Vitamin B12, 10 mg; Panthotenic acids, 5,000 mg; Folic acid, 500 mg; Biotin, 20 mg; Choline chloride 200 g; Manganese, 80 g; Zinc, 50 g; Iron, 20 g; Copper, 5 g; Iodine 12 g' Selenium, 200 mg; Cobalt, 200 mg.

Carcass and Haematological Indices

At the end of the experiment, three birds per replicate were randomly selected starved overnight and slaughtered by severing the jugular vein, the blood was put in labeled sterile universal bottles containing ethyl diamine tetra acetic acid (EDTA) powder as anticoagulant. The same birds were eviscerated for internal organs measurement. The percentage of packed cell volume (PCV) was determined by centrifugation of capillary tubes for 5minutes at 1200rpm, the haemoglobin content (HB) was determined by the methods of Jain [12]. The Red Blood Cells (RBC) was determined using the Hendricks fluid in an improved neubaur ruling counter chamber.

Statistical Analysis

Data collected were subjected to analysis of Variance. Differences between the treatment means were separated using Duncan's Multiple Range Test [13]. All statistical procedures were according to the methods of Steel and Torrie [14].

RESULTS AND DISCUSSION

The proximate composition nutritional factors of the test ingredient (table 1) fall within the range of values reported by Esonu et al., [15] for Hura crepitans meal. The raw seed meal is moderate in protein (24.80%) and crude fibre (11.82%) and very high in ether extract (39.45%). The protein and ether extract values were within the range of 23-28% crude protein and 38.95-51.24% of either extract reported by Esonu et al., [15], Olumide et al., [16]. The seed was also moderate in trypsin inhibitor (0.92mg/g), Tannin Cyanide (0.23mg/g) (0.51 mg/g),and saponin (0.33mg/g) when compared to other and seeds like mucuna bean, sovabeans as reported by Ukachukwu and Obioha [17].

There were significant (p<0.05) declines in the final live weight (FLW) and Average weight gain (AWG) as the level of Hura crepitans meal increased in the diets above 10%. The observed depression may be attributed to the tannin and phytates content of raw *Hura crepitans* meal which increased with increasing dietary levels. The presence of these anti- nutritional factors in feeds has been shown to reduce growth rate of broilers due to reduced protein absorption and specific amino acids utilization [18-20, 16]. However, the depression effect of these parameters from 10% inclusion levels of Raw *Hura crepitans* meal in broiler

diets contradict the level of inclusion of Hura crepitans meal in broiler diet where depression was only noticed from 15% inclusion levels as the report of Esonu *et al.*, [15]. These differences could be due to differences in nutrient content of the diet as well as feedstuffs composition of researchers' diets.

The feed consumption was equally significantly (p<0.05) influenced by inclusion level of raw $Hura\ crepitans$ meal and also for birds on T_4 (15%) and T_5 (20%) who showed significant (p<0.05) depression.

Hura crepitans meal is fibrous compared to soybean and maize, it is replacing, high level of it means highly fibre level, this according to savory and gentle should have increased intake to allow birds meet their dietary components other than energy but the reverse was the case. The contradiction could be attributed to the high energy density of the feed. raw Hura crepitans meal has high oil content (39.4%), increasing level of raw Hura crepitans meal in the feed means increase in energy density of feed, and birds are known to eat to meet its energy needs and once satisfied, intake is hampered [21].

Tannins are known to lower feed intake and reduced growth rate if their threshold levels are exceeded in a diet [22]. High level of raw *Hura crepitans* meal ranslates to high level of tannin. It is therefore likely that 15% and 20% inclusion of raw *Hura crepitans* meal has shut up the tannin above the threshold level of 0.5% as reported by Jansman [22].

The reduced feed intake may equally leads to reduce weight gain

Raw *Hura crepitans* meal inclusion similarly significantly (P<0.05) declined feed conversion ratio and protein efficiency at 15% and 20% *Hura crepitans* meal

Significant (p<0.05) depression in feed utilization and protein efficiency as the level of HSM increases in the diets agrees with Birk and Peri [23] who noted that phytates is capable of forming complexes with cations resulting in reduced availability of calcium, magnesium, potassium and copper and that with reduced availability of these nutrients and minerals, animals consuming diets with some probable level of phytates may not be able meet their nutrient requirements for tissue accretion hence the feed utilization is impaired [24].

Hura crepitans meal has reasonable level of phytates and fibre and when included in a high level will surely depress feed and protein utilization. The results of poor feed utilization is also in line with Ortiz et al., [25], who reported containing extract on feed gain ratio. Also high level of fibre according to

Onyeikegbulem [26] decreased the utilization of crude protein and other nutrients.

Results of nutrient digestibility showed a significant (p<0.05) decrease in crude protein, crude fibre and ether extract from 15% inclusion level of HSM. The observed low fibre and protein utilization from 15% raw *Hura crepitans* meal attributes to the high fibre contact of the feed at that level, according to Hedge *et al.*, [27]; Trait and Writ [28], fibre decreases the availability of nutrient by reducing the period of exposure of the food to the digestive enzymes and absorptive surfaces and this according to them is due to increased rate of passage induced by fibre which in turn affects the absorption of nutrients.

Also high level of oil in the feed limits the utilization of minerals and vitamins which could help in nutrient absorption [3].

Phytic acid lowers the bioavailability of minerals and inhibits enzymatic digestion of ingested proteins [29]. This could also be responsible for the poor nutrient retention when inclusion levels of raw Hura crepitans meal exceed 10% in the broiler diets.

Among the internal organs measured, only the liver showed significant differences. Percentage weight of liver was significantly (p<0.05) depressed as the level of raw *Hura crepitans* meal exceed 10%. This confirms the claim that liver may be the primary target

organ of anti-nutritional factors present in the raw seed meal. This results is in line with findings by [30-32] who observed birds fed with meal with anti-nutritional factors to have shrink liver, they all concluded that the primary target organ for anti-nutritional factor is the liver.

The abdominal fat deposit increases significantly as the level of raw *Hura crepitans* meal increased in the diet. This observation corresponds with the report of Nsa *et al.*, [3], who earlier reported that high fibre and high oil feed encourage abdominal fat deposit in broiler birds. HSM is of high fibre and oil when compared to the maize and soybean it is replacing in the diets.

Treatment diets 5 (20%HSM) showed significant (p<0.05) reduction of red blood cells (RBC) and packed cell volume (PVC) and hemoglobin (Hb) The haematological reduction suggests that trypsin inhibitor could be implicated. The raw sand box had earlier been found to have trypsin inhibitor [8] activity Ohoghobo *et al.*, [33] reported localization of trypsin inhibitor in a base soluble fraction of lima bean. They led the base soluble protein fraction to broilers and observed consistent reduction of RBC and HB in the birds. They concluded that this suggest, the direct involvement of trypsin inhibitor. However, the values of all the measured haematological indices were within limits for avian species [34].

Table-3: Performance characteristic and haematological indices of broiler birds fed diets containing raw Hura crepitans meal

Parameters inclusion level of raw Hura crepitans									
	0%	5%	10%	15%	20%	SEM			
Initial Weight	810.50	812.00	816.10	809.45	809.55	11.89			
Final live Weight	2980.16 ^a	3100.09a	2980.00a	2816.50 ^b	2440.00 ^c	9.99			
Weight gain(g/bird/day)	41.66 ^a	42.00a	40.91 ^a	34.68 ^b	27.14 ^c	2.65			
Feed intake(g/bird/day)	126.43a	128.94 ^a	126.61 ^a	114.90 ^b	101.00 ^b	7.22			
Feed gain(g of feed/g of Wt gain)	3.03°	3.02 ^c	3.09 ^c	3.40 ^b	3.72a	0.65			
Protein intake(g)	26.73 ^a	27.19 ^a	26.66a	24.17 ^b	21.26 ^c	1.18			
Mortality (%)	0.00	0.20	0.03	0.00	0.10	0.00			
Cost of 1kg feed consumed(N)	97.75 ^a	97.00 ^b	96.25°	95.50 ^d	94.69 ^e	10.16			
Cost of 1kg feed consumed/Weight gain(N/g)	296.65°	297.79°	297.88 ^c	316.41 ^b	352.73 ^a	22.17			
Feed cost savings (%)	-	-0.38	-0.41	-6.66	-18.90	0.00			
Haematological indices									
Packed cell volume (%)	39.10 ^a	38.92a	38.91a	37.05 ^a	33.66 ^b	5.88			
Hemoglobin(g/dl)	13.00 ^a	13.05 ^a	12.92ª	12.71 ^a	10.11 ^b	1.43			
Red blood cell(g/dl)	4.33	4.28a	4.29a	4.07	3.41b	0.83			
White blood cell(x103dl)	6.01	6.22	6.18	6.29	6.10	0.59			
Platelet(g/dl)	175.40	178.00	171.66	172.00	169.44	22.67			

Relative weights of organs (%) Dressing percentage 79.10 78.72 76.16 76.10 74.60 10.10 0.79 0.78 0.76 0.74 0.71 0.06 Kidney 0.22 0.23 0.27 0.27 Spleen 0.25 0.01 2.07a 1.85^b 1.76^b 0.32 Liver 2.10^{a} 2.05a Pancreas 0.33 0.31 0.31 0.31 0.30 0.04 0.59 0.60 0.62 0.62 0.65 0,02 Heart 0.71 0.72 0.71 0.74 0.74 0.06 Lungs Abdominal fat 0.61^{d} 0.78° 0.98^{b} 0.34^{e} 1.11^{a} 0,11 **Nutrient retention (%)** 81.49 Dry matter 81.65 81.32 81.20 81.01 11.20 Crude protein 78.41a 76.44a 76.02a 72.92^{b} 72.51^{b} 2.22 Crude fibre 37.46^{a} 37.66a 37.81a 34.30^b 30.05^{c} 1.90 65.72^b 71.65a 71.90a 72.17^a 71.40a 2.73 Ether extract Nitrogen free extract 80.11 80.20 80.23 80.44 80.72 3.18

Table-4: Organs weights and nutrient retention of broiler birds fed raw *Hura crepitans meal*

CONCLUSION

The result of this study revealed that HSM should not be included above 10% in broiler diets at can impaired performance. However, it is believed that with a good processing method to knock out the activity factors, a higher level might be tolerated and improvement in the growth performance parameters could be achieved in broiler birds' production.

REFERENCES

- Oladunjoye AO, Omogbemile D. Design and construction of two compartment freezing unit. 2003
- 2. Nsa EE, Ukachukwu SN, Akpan IA, Okon B, Effiong OO, Oko OO. Growth performance, internal organ development and hematological responses of broiler birds fed diets containing different thermal treated castor oil seed meal (Ricinus communis). Global Journal of Agricultural Sciences. 2010;9(2):27-34.
- Nsa EE, Akpan IA, Okon B, Anya MI. Performance of broiler finisher birds fed palm oil slurry as energy source. Nigerian Southeast Journal of Agricultural Economics and Extension, 2008; 8(1), 18-21.
- Oyebiyi OO, Farinu GO, Togun VA, Akinlade JA, Ajibola HO, Olaniyonu BI. Studies on growth and haematological attributes of weaner rabbits fed graded levels of sundried cassava peel-blood meal mixture. InProceedings of the 32nd Conference, Nigerian Society for Animal Production (NSAP). March 2007 Mar 18, 18-21.
- 5. Shonekam FO, Ajayi JO. The biochemical analysis of *Hura crepitance* (sand box) seed oil and meal. 2016. Globalacademicgroup.com
- 6. Nnaji C. Effect of different processing methods on some anti-nutritional compounds in sandbox (Hura crepitans) seed R. Agric. Tech. thesis. Redal University of Technology Oweri, Nigeria. 2010.
- 7. Oyeleke GO, Olayiwola OA, Latona DF. Chemical examination of sandbox (*Hura crepitans*) seeds:

- Proximate elements and fatty acid profile. Journal of Applied Chemistry, 2012; 1(2), 10-13.
- 8. Gbadamosi SO, Osungbade OR. Effects of cooking and fermentation on the antinutrients, total phenolic contents and antioxidant properties of sandbox (Hura crepitans) seeds. International Food Research Journal. 2017 Aug 1;24(4):1621-1627.
- Association of Official Analytical Chemists (AOAC). Association of Official Agricultural Chemist Inc. Official methods of analysis 15th edi. Assoc. Offic. Agric. Chem. Washington D. C. 1990.
- 10. Mega JA. Phytate: its chemistry, nutritional significance and methods of analysis. Journal of Agriculture and Food Chemistry, 2002; 30:1-9.
- 11. Liener IE. The photometric determination of the hemagglutinating activity of soyin and crude soybean extracts. Archives of biochemistry and biophysics. 1955 Jan 1;54(1):223-231.
- 12. Jain NC. Schalm's Vertinary haematology. 4th Edi., Lea and Febiger, Philadelphia, 1986; 149-162.
- 13. Duncan DB. Multiple range and multiple F tests. Biometrics. 1995 Mar 1;11(1):1-42.
- 14. Steel RG, Torrie JH. Principles and procedures of statistics, a biometrical approach. McGraw-Hill Kogakusha, Ltd.; 1980.
- Esonu BO, Ozeudu E, Emenalom OO, Nnaji C, Onyeikegbulem IK. Nutritional value of sandbox (Hura crepitans) seed meal for broiler finisher birds. Journal of Natural Sciences Research. 2014;4(23):95-9.
- 16. Fawale OS, Gbadamosi SO, Ige MM, Kadiri O. Effects of cooking and fermentation on the chemical composition, functional, and antinutritional properties of kariya (Hildergardia barteri) seeds. Food science & nutrition. 2017 Nov;5(6):1106-15.
- 17. Ukachukwu SN, Obioha FC. Effects of time duration of thermal treatments on the nutritive value of Mucuna cochinnensis. Journal of Applied. Chemistry and Agricultural Research, 1997; 4:39-

- 18. Douglas JH, Sullivan TW, Abdul-Kadir R, Rupnow JH. Influence of infrared (micronization) treatment on the nutritional value of corn and low-and high-tannin sorghum. Poultry Science. 1991 Jul 1;70(7):1534-1539.
- 19. Elkin RG, Rogler JC, Sullivan TW. Differential response of ducks and chicks to dietary sorghum tannins. Journal of the Science of Food and Agriculture. 1991;57(4):543-553.
- Olaposi RA, Oladipupo Q, Olumide S. Nutritional Physiology Chemical and functional properties of protein concentrate and isolate of newly developed Bambara groundnut (Vigna subterrenea L.) cultivars, Food Science and Nutrition, 2017; 6(1), 229-242.
- 21. Nsa EE, Okon BA, Kpan IA, Anya M, Woger GSI, Edet GD, Okereke CO, Juobi VI. Growth performance and economy of maize offal as a replacement for maize in broiler finisher diet. Proceedings of 14th Annual Conference of Animal Science Association of Nigeria (ASAN), Ogbomoso, 2009 September; 14-17.
- 22. Jansman AJ. Tannins in feedstuffs for simple-stomached animals. Nutrition research reviews. 1993 Jan;6(1):209-236.
- 23. Birk Y, Peri I. Phytates. In: *Toxic constituents of plants food stuffs*, Liener IE. (Ed). Academy Press New York. 1980; 161-182.
- 24. Fanimo AO, Adebayo AJ, Oduguwa OO, Biobaku WO. Feeding value of cashew testa for broiler chickens. Nigerian Journal of Animal Production. 2007;34(1):83-93.
- 25. Ortiz LT, Alzueta C, Trevino J, Castano M. Effects of faba bean tannins on the growth and histological structure of the intestinal tract and liver of chicks and rats. British Poultry Science. 1994 Dec 1;35(5):743-54.

- Onyeikejbulem IK. Studies on nutritional evaluation of raw and boiled sandbox (Hura crepitans) seed meal for broiler starter. B. Agric. Tech. Thesis. Federal University of Technology. Oweri Nigeria. 2011.
- 27. Hegde SN, Rolls BA, Turvey A, Coates ME. The effects on chicks of dietary fibre from different sources: a growth factor in wheat bran. British Journal of Nutrition. 1978 Jul;40(1):63-69.
- Brand AG. Writing and feelings: Checking our vital signs. Rhetoric review. 1990 Mar 1;8(2):290-308.
- Nolan KB, Duffin PA. Effect of phytates on mineral bioavcailability in vitro studies on Mg2+, Ca2+, Fe3+, Cu2+ and Zn2+ solubilities in the presence of phytates. Journal of Science, Food and Agriculture, 1987, 40:413-415.
- 30. Salgarkar S, Sohonie K. Haemagglutinins of field bean (Dolichos lablab). II. Effect of feeding field bean haemagglutinin A on rat growth. Indian journal of biochemistry. 1965 Sep;2(3):197-9.
- 31. Bassir O, Ikegwuonu F. The in vivo effects of phytohemagglutinins on ATPase and fumarase enzymes in the rat. Toxicon. 1975 Nov 1;13(5):371-4.
- 32. Ukachukwu SN. Effect of composite cassava meal with or without palm oil and/or methionine supplementation on broiler performance. Livestock Research for Rural Development, 2008; 20(4).
- Ologhobo AD. Nutritive values of some tropical (West African) legumes for poultry. Journal of Applied Animal Research. 1992 Dec 1;2(2):93-104
- 34. Fraser CM, Mays A. *The Merck Veterinary Manual: A Handbook of Diagnosis. Therapy and Diseases prevention and Control for the Veterinarian.* 6th edition. Merck and Co Inc. Ralway. New Jersey, USA. 1986, 1613-1614.