Research Article

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Effect of Jatropha curcas Supplemented Diet on Broilers

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Abstract: Several authors have shown that farmers in Africa feed their poultry with partial or total dietary supplement, especially, diet based on local and cheaper ingredients in order to maximize profit, this however may have opposite effects because some of these supplements are toxic to the birds leading to low productivity or high mortality among the birds. This study was therefore conducted to determine the effect of raw *Jatropha curcas* supplemented diet on broilers' health and growth using some biochemical parameters. A total of forty one-day old Arbor acres broilers were used for this study, they were acclimatize for four weeks during which medication and other managerial practice were administered to the birds as directed by the supplier before they were randomly allocated into four different experimental groups receiving different experimental diets containing 0, 4, 8 and 12% raw *Jatropha curcas* respectively. After four weeks of treatment with different experimental diets, blood samples were collected from the birds for biochemical analysis. The results showed that feeding broilers with raw *Jatropha curcas* supplemented feed resulted in elevation in liver biomarkers (AST, ALT, ALP ,Bilirubin) and kidney biomarkers (Urea, Creatinine). Also there was increase in serum cholesterol with a marked decrease in both serum protein and packed cell volume. From the result, it was obvious that raw *Jatropha curcas* supplemented diet is hepatotoxic, nephrotoxic and can affect blood circulation in the birds even in small quantity and should be detoxified before it can be used as a supplement or substitute in broiler's feed. **Keywords**: *Jatropha curcas*, supplement, broilers, toxic, biomarkers

INTRODUCTION

The common protein sources and other ingredients used in poultry feeds have become too expensive because of their high demand this had led to the evaluation of lesser known and underutilized crop seeds for their nutritive values. In this regards, *Jatropha curcas* seeds have been proposed as a cheap supplement to the conventional forage crops [1-3].

Jatropha curcas, а member of the Euphorbiaceae family is a drought resistant multipurpose tree of significant economic importance. The kernel has a protein content of 27-32 % by weight while the residue after oil extraction (fully defatted meal) has a relatively high protein content around 53-58% by weight [4]. This relatively high protein content of Jatropha curcas can be advantageous since this rich source of protein is not utilized by humans like commonly consumed food crops such as soy and wheat [4,5], however, at present the use of *Jatropha* in feed is limited owing to the presence of toxic and anti nutritional constituents[6].Several studies showed that the Jatropha curcas seeds are toxic to humans and animals due to the presence of phorbol esters and certain proteins [4, 6-9]. The biological effects of these compounds include tumors promotion, wide range of negative biochemical and cellular effects, alteration of cell morphology, induction of platelet aggregation and also serve as lymphocyte mitogens [10].

To maximize profit farmers have been feeding their poultry with partial or total dietary supplement, particularly diet based on local and cheaper ingredients [3, 11-17]. This however may have negative effects on the bird because some of these supplements are toxic to the animals leading to low productivity or high mortality among the birds. Therefore, it is necessary to investigate the toxicity of some of these supplements before they are used as supplement.

The purpose of this study was to evaluate the effects of raw *Jatropha curcas* incorporation into broiler's diets on some biochemical parameters in broiler in order to assess its toxicity

EXPERIMENTAL SECTION Sample Collection And Preparation

Jatropha curcas seeds were collected from Agwada area of Nasarawa state Nigeria and proper identification was carried out at the National Institute for Pharmaceutical Research and Development (NIPRD) located at Idu industrial area – Abuja. The seeds were dried and hand-cracked. The kernels obtained were milled using a mechanical grinder, air dried at room temperature and then packaged until when needed.

Birds Management and duration of the experiment

A total of 40 one- day old Arbor acres broiler chicks strain were purchased from Mararaba, Nasarawa state. They were brooded together for four weeks and fed with commercial broiler starter obtained from poultry feed vendor. All medications and required managerial practices were applied as at when due. After four weeks, the chicks were divided into their various experimental groups with each group receiving one of the experimental diets and clean drinking water adlibitum for a period of four weeks.

Commercial broiler diet

The commercial feed used was obtained from a reputable feed company and recompose into the desired experimental diet.

Table 1: Compositions of the commercial Feed

| Constituents | Percentage composition (%) |
|----------------------------|----------------------------|
| Cereals/grains | 25 |
| Vegetable protein | 13 |
| Premix (vitamins/minerals) | 9 |
| Essential Amino acids | 20 |
| Salt | 5 |
| Antioxidants | 9 |
| Anti-toxins | 9 |
| Prebiotic | 5 |
| Enzymes | 5 |
| Nutritional composition | |
| Crude protein | 20 |
| Crude fat | 10 |
| Crude fibre | 9 |
| Calcium | 1 |
| Available phosphorous | 0.45 |
| Metabolizable Energy | 2800kcal/kg |

SOURCE: Label on the feed as stated by the feed company

Experimental design

After four weeks of acclimatization, the birds were randomly allocated into four different dietary groups as shown below.

Group 1: Fed with normal commercial diet without *Jatropha curcas*.

Group 2: Fed with experimental diet containing 4% *Jatropha* curcas by composition.

Group 3: Fed with experimental diet containing 8% *Jatropha* curcas by composition.

Group 4: Fed with experimental diet containing 12% *Jatropha curcas* by composition.

| Composition | Group 1 | Group 2 | Group 3 | Group 4 |
|-------------------------|---------|---------|---------|---------|
| Commercial diet (%) | 100 | 96 | 92 | 88 |
| Raw Jatropha curcas (%) | 0 | 4 | 8 | 12 |
| Total (%) | 100 | 100 | 100 | 100 |

Table 2: Experimental diet composition

Physical and Clinical observations

The birds were keenly observed for any discharge, evidenced allergic reaction, behavioural changes, change in weight, mortality and any appearance of pathological condition

Analysis of raw Jatropha curcas

The moisture, protein, fat, ash, fiber and nitrogen free extract were analysed by the methods of AOAC[18]. Phytate was determined using the method descibed by Mohamed et al.[19] While Saponin and total oxalate were determined according to the methods of Pearson [20] and Adeniyi et al., 2012 respectively.

Collection of blood samples

5ml of blood samples were collected in duplicate from each bird. The first one was collected

into an anticoagulant bottle while the second one was collected into plain bottle without anticoagulant.

Haematological study

The blood samples in the anticoagulant (heparinized) bottles were used to determine the packed cell volume (PCV).

Serum biochemistry study

The blood sample in the plain bottles was allowed to clot for about two hour. The clotted blood was centrifuged at 3,500rpm for 30mins to recover the serum from the clotted blood. Serum was separated with sterile syringes and needles and stored frozen until when needed. The following biochemical analysis was performed on the sera samples: Serum aspartate amino transferase (AST) and alanine amino transferase (ALT) activities were estimated with the Randox reagent kit using 2, 4-dinitrophenylhydrazine as substrate according to the method described by Reitman and Frankel [22]. Urea, creatinine and total bilirubin concentrations were determined by the methods of Patton and Crouch [23]; Henry *et al.*[24] and Pearlman and Lee [25], respectively. Total cholesterol was measured by the procedure described by Allain *et al.*,[26]. Protein content was determined by the method of Lowry *et al.*[27] while Alkaline phosphatase was assessed as described by Principato et al.,[28]. All the assays were carried out at the Department of Biochemistry, Bingham University Karu Nasarawa state, Nigeria.

Statistical analysis

The data are expressed as mean \pm SD. Statistical analysis was performed using SPSS (Version 17). A level of p <0.05 was considered to be significant results

RESULTS AND DISCUSSION

The nutritional compositions of raw (undefatted) *Jatropha curcas* seeds are shown in tables 1. The moisture contents is lower than 10% moisture content limit recommended for storage stability of flours[29] which suggest that the seed flour will have a

long shelf life. The ash content (table1) of the J. curcas seed flour is an indication that it may have a reasonable quantity of mineral elements for building healthy body and proper function of the body tissues while the moderate fiber content means it will enhance easy movement of the bolus in the large intestine. The average crude protein obtained $(33.07 \pm 0.09\%)$ is higher than the value reported by Abou- Arab and Abu-Salem[30] and that of the seed of Jatropha gossipifolia (13.40) reported by Ogbobe and Akano[31]. This high crude protein content makes it a good source of protein and substitute or supplement for soybean and other protein rich legumes.

As expected high value was observed for the crude fat (34.01%) (Table1) which has been shown to contain most of the antinutritional factors especially phorbol. Relatively high carbohydrate level (20.29%) was also obtained. Crude fibre content (2.69%) (Table 1) is however lower than that reported by Abou- Arab and Abu-Salem [30] for raw *Jatropha curcas* but higher than 0.2% reported for soybean [32]. Crude fiber in diet consists mostly of plant polysaccharides that cannot be digested, it therefore increases stool bulk and decreases the time that waste materials spend in the gastrointestinal tract. [33, 34].

| Composition | Percentage (%) |
|-----------------------|-------------------|
| Moisture | 5.97 ± 0.014 |
| Ash | 6.76 ± 1.380 |
| Crude protein | 33.07 ± 0.090 |
| Crude fat | 34.01 ± 0.790 |
| Carbohydrates | 20.29 ± 0.160 |
| Crude fiber | 2.69 ± 0.860 |
| Nitrogen free extract | 5.20 ± 0.140 |

Table 3: Proximate composition of Jatropha curcas

All values are mean of triplicate determination \pm standard deviation.

Evaluation of antinutional content of raw *Jatropha curcas* showed that the saponin content (Table 4) is lower than 4700mg/100g reported by Makkar et al., [29] for Soybean meal. Saponin is linked with some negative effects on animals including reduction of palatability and intake of nutrient [35, 36]. The phytate content (Table 4) in the present study agrees with that of soybeans reported by Reddy and Pierson [37].

Phytates have been implicated in decreasing protein digestibility by forming complexes and also by interacting with enzymes such as trypsin and pepsin[37]. It also form complexes with divalent minerals thereby decreasing the bioavailability of these elements for absorption[38]. The high concentration of oxalate observed can affect calcium absorption.

| Table 4: Antihutritional factors in raw <i>Jatropha curcus</i> | | |
|--|-------------------|--|
| Anti-nutritional factors (mg/100g) | Content(mg/100g) | |
| Phytate | 48 ± 0.57 | |
| Saponin | 2500 ± 141.42 | |
| Oxalate | 16.5 ± 0.85 | |

Table 4: Antinutritional factors in raw Jatropha curcas

All values are mean of triplicate determination \pm standard deviation

Measurement of the activities of various enzymes in body fluids plays a significant role in disease investigation, diagnosis and detection of tissue damage. From the results (Table 5) there was an increase in the liver biomarker enzymes examined, this increase was dose dependent, that is, increase in the

quantity of *Jatropha curcas* in the feed resulted in increase in the level of these enzymes in the serum. The increase in serum liver enzymes (ALT, AST, and ALP) levels is an indicator of liver damage and cytotoxic effect of raw J. *curcas* seed on the liver cells leading to leakage of these enzymes from damaged hepatocytes cytosols into bloodstream[39,40,41]. ALT an enzyme produced in cytosol of hepatocytes of the liver is the most sensitive marker for liver than the remaining ones and it's serum level is an indication of hepatocellular damage that can therefore provide quantitative assessment of the degree of damage sustained by the liver [42].The trend of ALP observed gave an

indication that the hepatic capacity of the liver is grossly affected by *Jatropha curcas*[43]. Total bilirubin was also elevated in the serum of the broilers administered with *Jatropha curcas*; bilirubin is a conventional indicator of liver diseases and its elevation in the serum has been associated with hepatocellular damage and hepatic biliary tract obstruction. These results agree with previous reports by El Badwi et al.[44] and Samia et al[45] who reported the negative effect of *Jatropha curcas* inclusion in feed. The toxic effects were ascribed to the toxic substances in the oil of raw *Jatropha curcas* [44, 45].

| GROUPS | AST(U/I) | ALT(U/I) | ALP(U/I) | BILIRUBIN (mg/dl)) |
|------------|-----------------------------------|--------------------------|-----------------------|-----------------------|
| Group 1 | 72.55 <u>+</u> 5.98 ^a | 64.26±5.67 ^a | 33±11.00 ^a | 0.53 ± 0.08^{a} |
| Group 2 | 84.00 ± 20.65^{a} | 83.08 ± 7.52^{a} | 46.20 ± 18.70^{a} | 0.73 ± 0.17^{a} |
| Group 3 | 106.00 <u>+</u> 9.58 ^b | 242.90 ± 8.90^{b} | 70.40 ± 27.60^{b} | 1.40 ± 0.03^{b} |
| Group 4 | 106.17 <u>+</u> 9.64 ^b | 283.8±11.00 ^b | 79.2 ± 14.34^{b} | 1.61 ± 0.06^{b} |

Table 5: Effect of different graded level of raw Jatropha curcas in broilers feeds on hepatic function biomarkers

The values are mean \pm standard deviation of five observations. Values with different superscript in the same column are significantly different at p< 0.05.

The effect of *Jatropha curcas* on the kidney functions was assessed by the levels of serum creatinine and urea, as the levels of serum urea and creatinines are often regarded as reliable markers of renal function [46]. The significant elevation of creatinine and urea (Table 6) is a pointer to renal dysfunction in chickens given *Jatropha curcas*[47]. Creatinine is a break-down product of creatine. It is usually produced at a fairly constant rate by the body and filtered out of the blood by the kidneys. If the filtering capacity of the kidney is

deficient, creatinine blood level rises [48, 49]. Urea is the major end product of protein catabolism in animals and is the primary vehicle for removal of toxic ammonia from the body. It is primarily produced in the liver and excreted by the kidneys. In general, increased urea levels are associated with compromise in kidney function [46,50].Therefore, from the result the toxicants or antinutrional factors in raw *Jatropha curcas* can cause a damage to the kidney there by distorting renal function.

| GROUP | UREA(mg/dl)) | CREATININE(mg/dl)) |
|---------|----------------------|-----------------------|
| Group 1 | 30.83 <u>+</u> 4.52 | 0.45 <u>+</u> 0.07 |
| Group 2 | 42.08 ± 4.01^{a} | 0.51 <u>+</u> 0.06 |
| Group 3 | 58.33 ± 466^{a} | 1.580 ± 0.227^{a} |
| Group 4 | 70.00 ± 4.79^{a} | 2.882 ± 0.611^{a} |

The values are mean \pm standard deviation of five observations. a = significant difference at p<0.05 compare with the Control.

The administration of *Jatropha curcas* caused an increase in serum cholesterol and marked decrease in serum protein concentration (hypoproteinemia)[Table 7].The hypoproteinemia observed may be attributed to the direct toxic effect of phorbol esters leading to degeneration and necrosis of hepatocytes[39,51] which are considered to be the main site of protein synthesis[52]. The interaction of phorbol ester with protein kinase C (PKC) affects activities of several enzymes, including the enzymes involved in biosynthesis of protein, DNA, polyamines, cell differentiation processes, and gene expression[39,53].This may be responsible for the low levels of protein in the serum of the chicken fed with graded levels of *Jatropha curcas*. Reddy and Salunkhe[54], also attribute the decreases in total protein to inhiition of protein utilization in the broilers. Increase in serum cholesterol in broilers fed with graded level of *Jatropha curcas*, may be as a result of stimulation of lipolytic hormones action on the fat depots due to inhibition of insulin[55,56,57].This elevated blood cholesterol levels have been reported by Omage *et al.*[58] as the most important risk factor in heart disease which result result in the death of the

chicken.

 Table 7: Effect of different graded level of raw Jatropha curcas in broilers feeds on their serum protein and cholesterol

| GROUP | TOTAL PROTIEN (mg/dl) | CHOLESTEROL(mg/dl) |
|---------|-------------------------|-----------------------|
| Group 1 | 8211.20 <u>+</u> 226.10 | 55.23 <u>+</u> 17.96 |
| Group 2 | 6326.40 <u>+</u> 440.40 | 72.56 <u>+</u> 12.94 |
| Group 3 | 5120 <u>+</u> 33.94 | 119.37 <u>+</u> 24.87 |
| Group4 | 4524 <u>+</u> 369.20 | 143.14 <u>+</u> 11.91 |

The values are mean \pm standard deviation of five observations.

Table 8 shows the results of the packed cell volume of the broilers progressive decrease in packed cell volume was observed as the level of raw *Jatropha curcas* supplement increases. The decline in % PCV with an increase in raw *Jatropha curcas* in the diet indicates blood loss and destruction of erythrocytes, decrease in % PCV with an increase in raw *Jatropha curcas* all point to the development of anaemia in the broilers. Chivandi et al.[59] reported that the anaemia

could have been haemorrhagic as a result of blood loss through the gastrointestinal tract (GIT). Furthermore damage to the GIT was also reported to have resulted in maldigestion and malabsorption of nutrients required for erythropoiesis[59]. This was also in line with our observations that broilers fed with higher concentration of raw *Jatropha curcas* have stunted growth and looked very weak

Table 8: Effect of different graded level of raw Jatropha curcas in broilers feeds on their packed cell volume.

| GROUP | PACKED CELL VOLUME(PCV) % |
|---------|---------------------------|
| Group 1 | 28.60 + 2.07 |
| Group 2 | 24.60 + 2.41 |
| Group 3 | 19.60 + 1.14 |
| Group 4 | 19.00 + 1.58 |

The values are mean \pm standard deviation of five observations in percentage

FURTHER OBSERVATIONS

During the administration of the experimental diets the broilers fed with higher concentration of raw *Jatropha curcas*(8% and 12%) have stunted growth, looked very weak and their feathers sheds often. Also, mortality rate increased according to the percentage of raw *Jatropha curcas* meal ingested but no death was recorded in the control group.

CONCLUSION

From the results and observations, it was evident that raw *Jatropha curcas* supplemented feed is hepatotoxic, nephrotoxic and can affect blood circulation in the birds even in small quantity and should therefore be detoxified before it can be used as a supplement or substitute in animal feed.

REFERENCE

- 1. Srinivasan N, Perumal S and George F; Proximate composition and functional properties of raw and processed *Jatropha curcas* L. Kernel meal. International Journal of Research in Pharmaceutical and Biomedical Sciences, 2013; 4 (1):183-195.
- [Doumbia F; L'approvisionnement en intrants de la filière avicole moderne au Sénégal. Thèse Med. Vét., EISMV: Dakar, 2002: 27.
- Ayssiwede SB, Dieng HB, Chrysostome CAAM, Hane MB, Mankor A, Dahouda M, Houinato MR, Hornick JL and Missohou A;

Effects of *Moringa oleifera* (Lam.) Leaves Meal Incorporation in Diets on Growth Performances, Carcass Characteristics and Economics Results of Growing Indigenous Senegal Chickens. Pakistan Journal of Nutrition, 2011; 10 (12): 1132-1145.

- 4. Hamarneh AI, Heeres HJ, Broekhuis AA and Picchioni F; Extraction of *Jatropha curcas* proteins and application in polyketone-based wood adhesives. International Journal of Adhesion and Adhesives, 2012; 30: 615–625.
- Lestari D, Mulder W J and Sanders JPM; Jatropha seed protein functional properties for technical applications. Journal of Biochemical Engineering, 2011; 53: 297-304.
- Devappa RK, Swamylingappa B; Biochemical and nutritional evaluation of *Jatropha* protein isolate prepared by steam injection heating for reduction of toxic and antinutritional factors. Journal of Science Food and Agriculture, 2008; 88(5): 911–919.
- 7. Haas W and Mittelbach M; Detoxification experiments with the seed oil from *Jatropha curcas* L., Ind Crop Prod 2000; 12:111–118.
- 8. Makkar HPS, Francis G, and Becker K; Protein concentrate from *Jatropha curcas* screw-pressed seed cake and toxic and antinutritional factors in protein concentrate. J. Sci Food Agric, 2008; 88:1542-1548.

- 9. Mampane KJ, Joubert PH and Hay LT; *Jatropha curcas*: use as a traditional Tswana medicine and its role as a cause of acute poisoning. Phytotherapy Res, 1987; 1: 50-51.
- Azzaz NAE, El-Nisr NA, Elsharkawy EE and Elmotleb EA ; Chemical and Pathological Evaluation of *Jatropha curcas* Seed Meal Toxicity With or Without Heat and Chemical Treatment, Australian Journal of Basic and Applied Sciences, 2011; 5(12): 49-59.
- Buldgen A F, Detimmerman BS and Compere R; Etude des paramètres démographiques et zootechniques de la poule locale dans le basin arachidier sénégalais. Revue Elev. Méd. Vét. Pays Trop, 1992; 45: 341-347.
- Kondombo SR, Wakkel RPK, Slingerland M, Nianogo AJ and Verstegen MWA; Effects of local feedstuff supplementation on performance and nutritional status of village chickens during the end of the rainy season in Burkina Faso. Revue Elev. Méd. Vet. Pays Trop, 2003; 56: 199-204.
- 13. Rashid MM, Islam NM, Roy BC, Jakobsen K and Lauridsen C; Effect of dietary supplementation of energy and protein on production performance and egg quality of scavenging crossbred hens in rural areas under tropical conditions, Livest. Res. Rural Dev, 2004; 16 (8).
- 14. Riise JC, Permin A, Mcainsh CVand Frederiksen L, 2004; Elevage de la volaille villageoise. Un manuel technique sur la production avicole a petite echelle. RDAPE: Copenhague, Danemark, 2004: 103.
- Pousga S, Boly H, Linderberg JE and Ogle B, 2006. Effect of supplementation on feed intake and performance of confined and Scavenging crossbred growing chickens in Burkina Faso. Trop. Anim. Health Prod, 2006; 38: 323-331.
- 16. Halima H, Neser FWC, Tadelle D, De Kock A and Van Marle-Koster E; Studies on the growth performance of native chicken ecotypes and RIR chicken under improved management system in Northwest Ethiopia, Livest. Res. Rural Dev, 2007; 18 (6).
- 17. Kingori AM, Tuitoek JK, Muiruri HK, Wachira AM and Birech EK; Protein intake of growing indigenous chickens on free-range and their response to supplementation, Int. J. Poult. Sci, 2007; 6: 617-621.
- AOAC (2000). Official Methods of Analysis of the Association of Official Analytical Chemists International 17th Ed. Published by the Association of Official Analytical Chemists International, Suite 400, 2200 Wilson Boulevard, Arlington, Virginia 22201-3301. USA.
- 19. Mohamed AI, Perera RAJ and Hafez YS; (1986). New chromphore for phytic acid determination. Cereal Chem., 1986; 63:475.

- Pearson D (1976). The Chemical Analysis of Food. 7 th ed. Churchill Livingstone, Edinburg London 1976.
- 21. Adeniyi SA, Orjiekwe CL and Ehiagbonare JE Determination of alkaloids and oxalates in some selected food samples in Nigeria African Journal of Biotechnology ,2009; 8 (1), 110-112.
- 22. Reitman S and Frankel S; A colorimetric method for the determination of glutamic oxaloacetic and glutamic pyruvic transaminases. Am. J. Clinical Pathology, 1957; 28: 56-66.
- 23. Patton CJ and Crouch SR; Spectrophotometeric and kinetics investigation of the Berthelot reaction for determination of ammonia. Analytical Chemistry, 1977; 49: 464-469.
- 24. Henry RJ, Cannon DC and Winkelman JW; Clinical Chemistry Principles and Techniques, 11th Ed. Happer and Row Publishers, New York.1974:1629.
- 25. Pearlman FC and Lee RTY; Detection and measurement of total bilirubin in serum with use of surfactants as solubilizing agents. Clinical Chemistry, 1974; 20: 447–453.
- 26. Allain CC, Poon LS, Chan CS, Richmond W and Fu PC;Enzymatic determination of total cholesterol. Clinical Chemistry,1974; (20).1: 470-475.
- 27. Lowry OH, Rosebrough NJ, Farr AL and Randall RJ 1951. Protein measurements with the folin phenol reagent. J. Biol Chem.,1951; 193: 265-275.
- Principato GB, Asia MC, Talesa V, Rosi G, and Giovannini E; Characterization of the soluble alkaline phosphatase from hepatopancreas of Squilla mantis L. Comparative Biochemistry and Physiology,1985; 80B: 801–804.
- 29. Makkar HPS, Aderibigbe AO and Becker K; Comparative evaluation of nontoxic and toxic varieties of *Jatropha curcas* for chemical composition, digestibility, protein degradability and toxic factors, Food chem,1998; 62(2): 207-215.
- 30. Abou- Arab AA and Abu-Salem, MF; Nutritional quality of *Jatropha curcas* seeds and effect of some physical and chemical treatments on their anti-nutritional factors. African J. Food Sci, 2010;4 (3):93-103.
- Ogbobe O and Akano V; The physic-chemical properties of the seed and seed oil of *Jatropha* gossipifolia. Plant Food for Hum. Nutr, 1993; 43: 197-200.
- 32. Suarez FL, Springfield J, Furne JK, Lohrmann TT, Kerr PS, Levitt MD;Gas production in humans ingesting soybean flour derived from beans naturally low in oligosaccharides.

American Journal of Clinical Nutrition,1999; 69: 135-140.

- Eze SO and Ibe OJ; Effect of fermentation on the nutritive value of B. Eurycoma "Achi". Chemical Society of Nigeria, 2005: 30, 1-5.
- 34. Amaechi NC; Nutritive and anti-nutritive evaluation of wonderful Kola (Buccholzia coricea) Seeds. Pakistan Journal of Nutrition, 2009; 8(8): 1120-1122.
- 35. Makkar HPS, BeckerK, Sporer F and Wink M; Studies on nutritive potential and toxic constituents of different provenances of *Jatropha curcas*. Journal of Agriculture and Food Chemistry, 1997; 45: 3152–3157.
- 36. Fenwick GR, Price KR, Tsukamoto C, and Okubo K;Saponins. In Saponins in Toxic Substances in Crop Plants, D'Mello FJP, Duffus CM, Duffus JH (Ed). Cambridge: The Royal Society of Chemistry, 1991.
- Reddy NR and Pierson MD. Reduction in antinutrional and toxic components in plant foods by fermentation. Food Res. Int., 1994; 27: 281-290.
- Oboh G, Akindahunsi AA and Oshodi AA; Dynamics of Phytate-Zn balance of Fungi Fermented Cassava products (Flour and Gari). Plants Food for Human Nutrition, 2003; 58, 1-7.
- 39. Nabil AEA, Neveen AE, Elsharkawy EE and Elmotleb EA;Chemical and Pathological Evaluation of *Jatropha curcas* Seed Meal Toxicity With or Without Heat and Chemical Treatment .Australian Journal of Basic and Applied Sciences,2011; 5(12): 49-59.
- Tietz NW Text;book of Clinical Chemistry .2nd ed. W. B. Sonuners Co.philadelphia, 1994;851-860.
- 41. Philip GH, Reddy PM, Sridevi G 1995. Cypermethrin induced in vivo alterations in the carbohydrate of 34 freshwater fish Labeo rohita. *Ecotoxicol. Environ. Safety*, 1995; 31: 173-178.
- 42. Aniagu SO, Nwinyi FC, Akumka DD, Ajoku GA, Dzarma S, Izebe KS, Ditse M, Nwaneri PEC, Wambebe C and Gamaniel K; Toxicity studies in Rats fed nature cure bitters. Afr. J. Biotechnol.,2004; 4(1): 72-78.
- 43. Kaneko JJ; Clinical Biochemistry of Domestic Animals. Academic Press, San Diego, CA;1989.
- 44. El Badwi, S.M.A., H.M. Mousa, S.E.I. Adam, and H.J. Hapke; Response of Brown Hisex chicks to low levels of *Jatropha curcas*, *Ricinus communis* or their mixture. Vet. Hum. Toxicol. 34: 304-306.
- 45. Samia MA, El Badwi SMA, Adam SEI, and Hapke HJ;mToxic effects of low levels of dietary *Jatropha curcas* seed on Brown Hisex chicks. Vet. Hum. Toxicol.,1992; 34: 112-115.

- Ojo RJ, Segilola LI, Ogundele OM, Akintayo CO and Seriki S;Biochemical evaluation of lima beans (*phaseolus lunatus*)in alloxan induced diabetic rats ARPN Journal of Agricultural and Biological Science 2013; 8,(4):302 -309.
- 47. Garba SH, Adelaiye AB, Mshelia LY; Histopathological and biochemical changes in the rats' kidney following exposure to a pyrethroid based mosquito coil. J. Appl. Sci. Res., 2007 3: 1788-1793.
- 48. Nwanjo HU,Okafor MC,Oze, GO; Changes in biochemical parameters of kidney function in rats co-administered with chloroquine and aspirin. J. Clin. Sci., 2005; 23: 10-12.
- 49. Finco DR; "Kidney function," in *Clinical Biochemistry of Domestic Animals*, Kaneko JJ, J. Harvey JW, and Bruss ML, Eds., Harcourt Brace and Company Asia PTE. Limited, Singapore, 5th edition, 1997; 441-484.
- Sumiati A S, Hidayah LN, and Santoso WB; Toxicity of *Jatropha curcas* L. meal toxins on Broilers. Proceeding of Seminar AINI (Indonesian association of Nutrition and Feed science) VI, July 26-27, 2007, 195-201.
- 51. Aregheore EM, Becker K and Makkar HPS; Detoxification of a toxic variety of *Jatropha curcas* using heat and chemical treatments, and preliminary nutritional evaluation with rats. S. Pac. J. Nat. Sci.,2003 21: 5-56.
- 52. Eisenbarth G.S; Carbohydrate and protein metabolism .N. Engl. J. Med, 1986;1314-1360.
- Goel G, Makkar HPS, Francis G and Becker K; Phorbol esters: structure, occurrence and biological activity. Int. J. Toxicol., 2007;26: 279-288
- 54. Reddy NR and Salunkhe DK; Phytates in legumes and cereals reviews Adv. Food Res., 1982; 28: 1-8.
- 55. Ojo R J, Memudu AE, Akintayo CO and Akpan IS; Effects of Pre-induction Administration of Allium *Sativum* on SomeBiochemical Parameters in Alloxan Induced Diabetic Rats Research Journal of Applied Sciences, Engineering and Technology 2012; 4(23): 5129-5135.
- 56. Goodman LS and A. Gilman A; The Pharmacological Basis of Therapeutics. Macmillan, New York, 1985;1490-1510.
- 57.]Liu PS, Lin MK.; Biphasic effects of chromium compounds on catecholamine secretion from bovine adrenal medullary cells. Toxicology; 1997; 117 (1):45- 53.
- 58. Omage JJ, Umar IA and Bawa GS; Effect of sesame (*Sesamum indicum* L.) seed oil on blood and liver lipid/cholesterol levels of rats fed a high fat diet. Nigeria Journal of Experiment and Applied Biology, 2002; 3: 125-129.

59. Chivandi E, Erlwanger KH, Makuza SM, Read JS and Mtimuni JP; Effects of Dietary *Jatropha curcas* Meal on Percent Packed Cell Volume, Serum Glucose, Cholesterol and Triglyceride Concentration and Alpha-Amylase Activity of Weaned Fattening Pigs. Research Journal of Animal and Veterinary Sciences, 2006;1(1):18-24.