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# **Original Research Article**

# Effects of the ethanolic extract of *Piliostigma reticulatum* DC (Horscht) stem bark on biochemical parameters of albino rat wistar

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**Abstract:** *Piliostigma reticulatum* is a medicinal plant used in traditional treatment of digestive disorders such as diarrhoea in Côte d' Ivoire. To verify its safety, the ethanolic extract of the stem bark is orally administered for 28 days in three groups of rats at doses of 250, 500 and 1000 mg / kg body weight. The control group received distilled water. The blood collected every week is used to assay biochemical parameters. At all doses, the ethanolic extract of the stem bark of *Piliostigma reticulatum* did not affect the serum values of transaminases, urea, total protein, triglycerides and bilirubin values compared to the control group. In return, it resulted in all dose levels, an increase in serum creatinine in the second week and an increase in serum cholesterol in a dose of 1000 mg / kg body weight at the first and fourth week. Our sample resulted in the fourth week, hypoglycemia in the group treated at 1000 mg / kg body weight. All these observed effects are reversible. Furthermore, it does not cause delayed toxicity.

Keywords: Biochemical parameters, ethanolic extract, Piliostigma reticulatum, rat, reversible, safety.

## **INTRODUCTION**

The use of medicinal plants for health care is a practice that is gaining popularity worldwide. With this in mind, WHO recommends the use of herbal medicine whose safety, efficacy and quality are guaranteed [25]. Piliostigma reticulatum is one of the medicinal plants of the ivorian rich flora [2], used for the treatment of several diseases such as colic, hemorrhoids, especially diarrhoea [31, 9]. Piliostigma reticulatum is a tree of 8 to 10 m high, bole rarely straight, sometimes bushy rejection by strain, with a rounded, bushy top. Its bark is deeply fissured and cracked, sometimes ferruginous gray with a pink fibrous slash turning brown. The leaves are alternate, leathery, couplets and hairless beneath. They are heavily lobed with rounded lobes or corner. The fruit is a woody pod, flat, hairless, sometimes twisted and cracked [5]. For the traditional treatment of diarrhoea, alcohol macerated of Piliostigma reticulatum stem bark is used in beverage for two or three days [9]. Pharmacological studies on this plant have shown that the leaves of *Piliostigma reticulatum* have antibacterial and antimicrobial properties [3]. The sedative and anticonvulsant properties of leaves were also highlighted [8]. With an estimated  $LD_{50}$  more than 5000 mg / kg of body weight, total ethanolic extract of stem bark of Piliostigma reticulatum reduced diarrhoeic feces at doses of 250, 500 and 1000 mg / kg body weight [9]. These authors showed that this sample to 500 and 1000 mg / kg body weight also inhibits the

gastro intestinal motility. Despite these multiple biological activities, safety-related studies have not been conducted. The aim of this study is to evaluate the subacute toxicity of total ethanolic extract of stem barks of *Piliostigma reticulatum* through its effects on some biochemical parameters.

#### MATERIAL AND METHODS Plant material

*Piliostigma reticulatum* stems bark were collected in January 2015 in Kadjabo in the township of Dimbokro located about 240 km from Abidjan. The identification was confirmed by the National Centre for Floristic of Abidjan where a sample is deposited under number 18033.

## Animal

Rats of the species *Ratus norvegicus*, wistar strain aged 4 to 6 weeks, weighing between 97 and 108 g were used for the test. They come from our pet store. All animals were subjected to a temperature of  $25 \pm 2^{\circ}$  C with alternating 12 hours of light and 12 hours dark. They were fed pellets of FACI<sup>®</sup> and were given tap water ad libitum in bottles without discontinuity.

## Preparation of total ethanolic extract of the stem bark of *Piliostigma reticulatum*

The stem bark of *Piliostigma reticulatum* are dried in the laboratory at a temperature of  $25 \pm 2 \degree C$  for two weeks and pulverized using a mixer brand

RETSH type SM 100. Fifty grams of this powder are soaked in 1L of ethanol solution (96°) / water (80:20) for 24 hours with magnetic stirring. The extract is filtered through cotton wool and then on paper whattman's No. 1. The filtrate was evaporated under vacuum using a rotary evaporator type BUCCHI R 110 / NKE 6540/2. The extracts were introduced in an oven at 45°C for 48 hours to obtain powders that constitute the total ethanolic extract of *Piliostigma reticulatum* stem bark [9].

#### Study of subacute toxicity

The study of subacute toxicity is conducted according to the OECD 407 guideline [24]. Sixty rats were divided into six groups comprising four tests groups and two control groups. Each group consisted of 10 rats with 5 males and 5 females. Three doses were selected based on the work of Dosso et al.[9]. Doses 250, 500 and 1000 mg / kg body weight (bw) are administered to groups respectively 2, 3 and 4. Group 1 was received distilled water. The animals are individually marked and dosed orally an equal volume of vehicle to 2 ml / 100 g bw. The groups 5 and 6 are satellite groups receiving respectively distilled water and the ethanolic extract of Piliostigma reticulatum at a dose of 1000 mg / kg bw. The latter two items were used to study the reversibility, persistence, or delayed occurrence of toxic effects for 14 days after stopping treatment.

#### **Blood samples**

The day of sampling, the animals are fasted from 10 p.m. to 7:00 in the morning without food but with water at will [24]. The morning they were anesthetized with ether and blood is collected by the technique of incising the tip of the tail which is previously disinfected with alcohol 96  $^{\circ}$  [18]. Blood is collected into dry tubes. These samples are taken in all rats one day prior to the administration of the extract, and weekly in the first four groups. Fourteen days after cessation of treatment, blood in the satellite groups of rats is taken.

#### **Blood biochemical examinations**

The glycaemia is determined directly from whole blood using a glucometer brand Accu-Chek<sup>®</sup> (Roche Diagnostics) according to the glucose oxidase method [30]. The blood contained in each dry tube was centrifuged at 3000 revolutions / minute for 5 minutes and the serum obtained was used for the measurement of other biochemical parameters. Glutamate pyruvate transaminase (GPT) and oxaloacetates (GOT) are determined by the kinetic method [13], total cholesterol and urea by the enzyme method [4, 14, 29]. Creatinine [10], total protein and triglycerides [11] were determined by the colorimetric method. Bilirubin is measured by the diazo method [21].

### Statistic analysis

Graph pad software version 5.01 was used for processing the data obtained. Values are presented as the averages followed by the standard error mean (SEM). The comparisons of means are made relative to the control, due to the repeated measures ANOVA with mixed model, followed by Bonferroni post hoc test for a 95% confidence interval. The differences are significant if the p value is less than 0.05 [15].

## RESULTS

Prior to administration of the extract, the values obtained on day 0 for all the studied parameters of the different groups were nearly equal to those from the control group (Figure 1 to 3 and Table I to III). Similarly, glucose and cholesterol levels did not significantly change known to 250 and 500 mg / kg b.w. during the experiment. But, after four weeks of administration of the extract, blood glucose treated rats at the dose of 1000 mg / kg b.w. decreased significantly (p <0.05) compared to the control group (Figure 1). Moreover, cholesterol of treated rats at the dose of 1000 mg / kg b.w. increased significantly (p <0.05) to the first and fourth week (Figure 2). Throughout the period of administration of the ethanol extract of the stem bark of Piliostigma reticulatum, serum triglycerides in all treated groups did not undergo major changes compared to the control group. As to creatinine, a very significant increase in serum (p < 0.01) was observed at the second week at all doses studied (Figure 3). However, any disturbance in serum urea and total protein was observed in all treated groups compared to control group. Also, the level of GOT, GPT and bilirubin in the serum of treated groups were not significantly different when compared to the values of the control group (Table II).

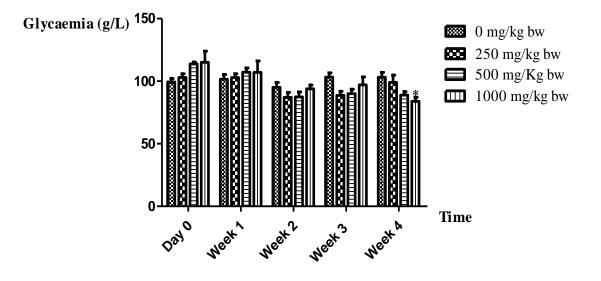


Figure 1: Effect of ethanolic extract of Piliostigma reticulatum stem bark on glycaemia,

n=10, \*: Statistically significant difference compared to the group 0 mg/kg bw (p < 0.05).

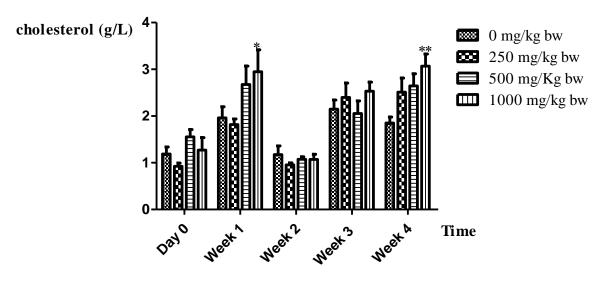


Figure 2: Effect of ethanolic extract of Piliostigma reticulatum stem bark on cholesterol,

n=10, \*: statistically significant difference compared to the group 0 mg/kg bw (p <0.05),\*\* statistically highly significant difference compared to the group 0 mg/kg bw (p <0.01).

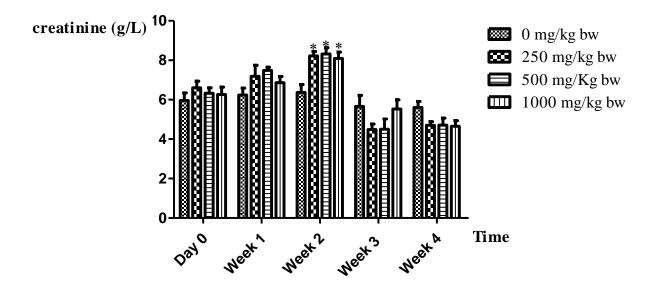


Figure 3: Effect of ethanolic extract of Piliostigma reticulatum stem bark on creatinine,

n=10, \*: Statistically significant difference compared to the group 0 mg/kg bw (p < 0.05).

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	Dose	D 0	w 1	w 2	w 3	w4
	(mg/kg					
	<b>b.w.</b> )					
	0	$0.8188 \pm 0.1387$	$0.8680 \pm 0.1346$	$0.7650 \pm 0.01688$	$0.6350 \pm 0.09459$	$0.6277 \pm 0.09765$
	250	0.7446±0.09611	0.8000±0.09517	$0.8050 \pm 0.05683$	0.7020±0.07912	$0.7015 \pm 0.09179$
Triglycerides	500	0.8326±0.1193	0.9130±0.1491	$0.6980 \pm 0.04742$	0.6950±0.04362	$0.6853 \pm 0.05266$
(g/L)	1000	0.7797±0.04363	$0.8490 \pm 0.05630$	$0.7690 \pm 0.03860$	$0.5760 \pm 0.09432$	$0.5529 \pm 0.08076$
	0	78.32±4.429	81.32±4.300	88.25±5.291	82.78±4.246	82.96±5.337
Total	250	86.26±6.178	88.18±6.131	81.59±9.537	89.82±5.726	87.06±4.917
proteins	500	87.79±4.369	87.33±5.370	88.63±5.119	87.95±4.588	85.71±3.536
(g/L)	1000	93.49±6.419	92.07±7.020	89.84±7.458	91.53±6.931	95.11±6.799
	0	2.589±0.1769	2.500±0.1905	2.664±0.2047	2.700±0.1606	2.858±0.1845
	250	2.713±0.1876	2.450±0.1364	2.938±0.1488	2.778±0.3403	3.116±0.3094
Urea (g/L)						
.3 /	500	2.682±0.1237	2.460±0.1300	2.910±0.2173	2.710±0.3455	3.294±0.3830
	1000	2.660±0.3159I	2.700±0.2366	2.841±0.2480	2.800±0.3091	3.522±0.2643

 Table I: Effect of the ethanolic extract of *Piliostigma reticulatum* on biochemical parameters of the liver and kidneys.

n = 10 for each week, testing groups are compared to dose 0 mg / kg bw; D 0 = 1 day before feedings; w1, w2, w3, w4 respectively first

, 2nd, 3rd and 4th weeks; threshold of significance  $\alpha=0.05.$ 

Table II Effect of the ethanol extract of <i>Piliostigma reticulatum</i> on biochemical markers of liver						
	Doses	D 0	w 1	w 2	w 3	w4
	mg/kg					
	b.w.					
	0	9.998±0.5123	11.15±0.6165	12.74±0.4383	12.59±0.8479	14.35±0.8131
GPT	250	9.111±1.505	11.34±1.515	12.29±0.4116	11.57±2.097	13.78±0.5872
(UI/L)	500	11.83±0.4488	13.33±0.6655	13.54±0.9157	15.00±1.057	15.09±0.9220
	1000	10.84±1.342	12.25±1.630	12.71±0.9421	13.73±1.985	14.34±1.307
	0	31.56±1.023	30.34±1.505	31.52±1.533	30.74±1.621	30.07±1.425
GOT	250	30.31±1.626	32.77±2.627	28.61±3.883	32.62±2.926	33.97±2.209
(UI/L)	500	25.23±1.873	31.89±4.019	33.26±1.668	36.38±3.157	37.17±2.634
	1000	31.68±2.204	33.83±3.753	32.52±1.184	33.98±2.974	38.48±2.060
	0	3.086±0.4222	2.601±0.3283	2.585±0.3056	2.567±0.3657	2.876±0.3609
Total	250	4.283±0.6070	3.589±0.5340	3.649±0.5660	3.595±0.5681	3.958±0.5770
Bilirubin	500	3.013±0.2607	2.497±0.2500	2.556±0.2438	2.435±0.1959	2.797±0.2628
(g/L)	1000	3.716±0.3756	3.027±0.2846	3.160±0.3003	2.931±0.2492	3.486±0.3732
	0	1.064±0.1337	0.8900±0.1167	0.9000±01137	0.8840±0.1194	0.9857±0.1271
Direct	250	1.491±0.2227	1.248±0.1880	1.253±0.1815	1.240±0.1906	1.365±0.1932
Bilirubin	500	1.042±0.09929	0.8602±007203	0.8710±0.07131	0.8494±0.06661	0.9579±0.08702
(g/L)	1000	1.275±0.1306	1.053±0.09904	1.066±0.09643	1.039±0.08988	1.178±0.1247

n = 10 for each week, testing groups are compared to dose 0 mg / kg bw; D 0 = 1 day before feedings; w 1, w 2, w 3, w 4 respectively 1rst, 2nd, 3rd and 4th weeks; threshold of significance  $\alpha = 0.05$ .

After discontinuation of the administration of the ethanolic extract of the stem bark of *Piliostigma reticulatum*, no significant change of the parameters of the treated rats were observed compared to the control group (**Table III**). Slight variations were noted at the end of the experiment at a dose of 1000 mg / kg body weight all disappeared after discontinuation of treatment. There was a decrease in blood glucose, increased serum cholesterol and creatinine.

Table III: Effects of ethanolic extract of <i>Piliostigma reticulatum</i> on biochemical parameters of satellite rats.
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	Dose mg/kg bw	D0	w6
Glycaemia (g/L)	0	$104.8 \pm 4.284$	$102.3 \pm 3.865$
	1000	$108.3 \pm 3.413$	$99.40 \pm 14.43$
Cholestérol	0	$1.466 \pm 0.4440$	$1.539 \pm 0.4962$
(g/L)	1000	$1.244 \pm 0.2325$	$1.121 \pm 0.2185$
Triglycerides	0	$0.6568 \pm 0.07483$	$1.264 \pm 0.1703$
(g/L)	1000	$0.7210 \pm 0.07223$	$2.071 \pm 0.4238$
Creatinine (g/L)	0	$6.232 \pm 0.2778$	$5.619 \pm 0.3867$
	1000	$5.413 \pm 0.2971$	$5.258 \pm 0.2222$
Urea	0	$2.633 \pm 0.3073$	$2.900 \pm 0.3966$
(g/L)	1000	$2.561 \pm 0.1776$	$3.307 \pm 0.3046$
<b>Total Proteins</b>	0	87.64 ± 6.721	83.76 ± 5.300
(g/L)	1000	81.15 ± 7.782	$77.72 \pm 7.429$
TGP (UI/L)	0	$11.45 \pm 1.640$	$12.85 \pm 1.829$
	1000	$12.74 \pm 1.969$	$14.15 \pm 2.108$
TGO	0	$31.29 \pm 2.205$	33.05 ± 3.105
(UI/L)	1000	$29.50 \pm 2.261$	34.31 ± 4.162
Total Bilirubin	0	$2.293 \pm 0.2856$	$2.598 \pm 0.4072$
(g/L)	1000	$2.747 \pm 0.3500$	$3.134 \pm 0.3411$
<b>Direct Bilirubin</b>	0	$0.8201 \pm 0.1237$	$0.9066 \pm 0.1355$
(g/L)	1000	$0.9815 \pm 0.1321$	$1.063 \pm 0.1193$

n = 10 for each week, testing group is compared to dose 0 mg / kg bw; D 0 = 1 day before feedings, S 6 = 6th weeks; threshold of significance  $\alpha = 0.05$ 

## DISCUSSION

Prior to administration of the ethanolic extract of *Piliostigma reticulatum* stem bark, all rats used for the experiment were in the same physiological

condition for the determination of biochemical parameters revealed identical values. During the experiment, serum creatinine increased in all treated rats at the second week. This result is similar to that of other authors who have shown an increase in serum creatinine in rats treated with the aqueous extract of Spondias mombin stem bark at doses 500 and 1000 mg / kg bw [22]. Creatinine and urea are markers of renal function. Thus, an increase of their rate reflects renal dysfunction [20]. It therefore appears signs of nephrotoxicity after two weeks of treatment. However, it seems that this is transitory nephrotoxicity because she had gone to the third and fourth week of the experiment. Transaminases reflect liver function, particularly glutamate pyruvate transaminase [26]. An increase of their rate indicates liver damage [16]. During the period of administration of the extract, serum transaminases of the treated groups were unchanged implying that the ethanol extract of the stem bark of Piliostigma reticulatum did not affect liver function. Previous works in our own have led to different results [12]. It has been observed an increase in serum GPT following repeated administration of the aqueous extract of Allium sativum to rats at a dose of 4800 mg / kg bw. They GOT serum level also provides information on the condition of skeletal and cardiac muscles [27]. Therefore, our findings suggest that the ethanol extract of the stem bark of Piliostigma reticulatum does not result in myocardial or skeletal muscles injuries of rats. This result is similar to that achieved by Koné et al. in their work on the subacute toxicity of the aqueous extract of Sacoglottis gabonensis stem bark in rats [17]. Bilirubin comes from the conversion of the heme released from the erythrocytes [28]. It may be in free or conjugated form. High bilirubin levels are due either to an inability of the kidneys to remove excess bilirubin or to a failure of the liver to combine effectively to its excretion [6]. Serum bilirubin levels did not change during the study, which supports the hypothesis that the extract tested would have no effect on the liver. This result is not consistent with that obtained in other experiments [23]. These authors showed with the aqueous extract of Cochlospermum planchonii rhizomes at a dose of 50 mg / kg bw, decreased bilirubin during an administration period between 1 and 5 days, followed by an increase between 10 and 15 days. A drop in proteinemia indicate a malfunction of microtubule system of hepatocytes [28]. The ethanol extract of Piliostigma reticulatum stem bark did not change the total protein. Our extract would therefore have no effect on the metabolism of proteins, particularly the microtubule system of rat hepatocytes. Increased levels of triglycerides and cholesterol promote arteriosclerosis which in turn is the cause of the majority of myocardial infarction [19]. Regarding the lipid profile, our extract resulted in an increase of the cholesterol to the first and fourth weeks for the dose of 1000 mg / kg bw but was without effect on triglycerides. Its effect is therefore directed specifically on the metabolism of cholesterol. This fact differs from that observed in the work of another authors [7]. They did not observed any variation in cholesterol levels with the aqueous extract of Elaeocarpus grandiflorus. In the fourth week, the

decline in blood glucose of rats at a dose of 1000 mg / kg bw suggests that the extract disrupt glucose metabolism in the fourth week. Results different from ours were obtained by administering about 28 days the aqueous extract of the stem bark of Sacoglottis gabonensis in rats [17]. Indeed, in their study, at doses of 3.5, 35 and 350 mg / kg bw, blood sugar has not changed. After stopping treatment, no change was observed for all studied parameters. This assumes that the ethanolic extract of the stem bark of *Piliostigma* reticulatum does not cause delayed toxicity. Furthermore, the toxic effects found during the experiment are reversible.

# CONCLUSION

This study found that the ethanol extract of the stem bark of *Piliostigma reticulatum* administered orally over 28 days, caused renal toxicity and a transient disturbance of lipid and carbohydrate metabolism. This extract is yet well tolerated by the body. However, the use of the ethanolic extract of *Piliostigma reticulatum* stem bark over a long period must be done with great caution since at high doses, it may cause side effects in humans. Histopathological analysis would be required to provide additional informations to those obtained in this study.

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## REFERENCES

- Adeoye BA, Oyedepo OO; Toxicity of Erythrophleum guineense stem bark: role of alkaloidal fraction. African Journal of Traditional Complementary and Alternative Medicine, 2004; 1: 45-54.
- Aké-Assi L; Rapport sur le colloque international, sur la médecine traditionnelle africaine à Abidjan. Revue Médicinale Traditionnelle, 1991; 4(2): 203-204.
- Akinsinde K, Olukoya D; Vibriocidal activities of some local herbs. Journal of .Diarrhoeal Diseases Research, 1995; 13(2): 127-136.
- Allain CC, Poon LS, Chan CSG, Richmond W, Fu PC; Enzymatic determination of total serum cholesterol. Clinical Chimistry, 1974; 20(4): 470-475.
- Arbonnier M; Arbres, arbustes et lianes des zones sèches de l'Afrique de l'Ouest. Edition 2, CIRAD, MNHN, Paris, 2002: 574.
- Bosma PJ; Inherited disorders of bilirubin metabolism. Journal of Hepatology, 2003; 38 (1): 107-117.

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- 7. Bualee C, Ounaroon A, Jeenapongsa R; Antidiabetic and long term effects of *Elaeocarpus* grandiflorus, Nar.Univ. J., 2007; 15(1): 17-28
- 8. Bum EN, Taiwe GS, Nkainsa LA, Moto FCO, Etet PS, Hiana IR *et al.;* Validation of anticonvulsant and sedative activity of six medicinal plants, Epilepsy behavior, 2009; 14(3): 454-462.
- Dosso K, N'guessan BB, Bidie AP, Gnangoran BN, Méité S, N'guessan D *et al.*; Antidiarrhoeal activity of an ethanol extract of the stem bark of *Piliostigma reticulatum* (Caesalpiniaceae) in rats. African Journal of Traditional Complementary and Alternative Medecine, 2012; 9(2): 242-249.
- Fabiny DL, Ertingshausen G; Automated reactionrate method for determination of serum creatinine with CentrifiChem. Clinical Chimistry, 1971; 17: 696-700.
- 11. Fossati P, Prencipe L; Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen peroxide. Clinical Chimistry, 1982; 28: 2077-2080.
- 12. Gatsing D, Aliyu R, Kuiate JR, Garba IH, Jaryum KH, Tedongmo N *et al.*; Toxicological evaluation of the aqueous extract of *Allium sativum* bulbs on laboratory mice and rats, Cameroonian Journal of Experimental Biology., 2005; 1(1): 39-45.
- Gella FJ, Olivella T, Cruz PM, Moreno R, Durban R, Gomez JA; A simple procedure for routine determination of aspartate aminotrans-ferase and alanine aminotransferase with pyridoxal phosphate. Clinica Chimica Acta, 1985; 153: 241-247.
- Gutmann I, Bergmeyer HU; Methods of enzymatic Analysis, ed Bergmeyer HU, Academic Press, NY. 4: 1794-1798.
- 15. Graph pad; Prism 5 for windows, Version 5.01. Graphpad Software Inc, 2007.
- Kew C; Serum aminotransferase concentration as evidence of hepatocellular damage. The Lancet, 2000; 355: 591-592.
- 17. Koné M, Bléyéré NM, Yapo AP, Vangah MO, Ehile EE; Evaluation de la toxicité d'un extrait aqueux de *Sacoglottis gabonensis* (Baille) Urban (Humiriaceae) chez les rongeurs, une plante utilisée dans le traitement de l'ulcère de Buruli en Côte d'Ivoire. International Journal of Biological and Chemical Sciences, 2009; 3(6): 1286-1296.
- Kraus AL; Research methodology, in The Laboratory Rat. Academic Press, New York, 1980: 1–30.
- Kris–Etherton PM, Nicolosi RJ; Trans fatty acids and coronary heart disease risk. Interntional Life Sciences Institute Press, Washington D.C, 1995: 1-24.
- 20. Levey AS, Miller WG, Greenberg N; Definition and classification of chronic kidney disease: a position statement from kidney disease improving global outcomes. Kidney International, 2005; 67: 2089-2100.

- 21. Malloy HT, Evelyn KA; The determination of bilirubin with the photoelectric colorimeter. Journal of Biological Chemistry, 1937; 119: 481-490.
- Moussa G, Koné M, Bleyéré NM, Yao KE, Yapo AP; Effect of total aqueous stem bark extract of *Spondias mombin* L. on some biochemical and anthropometric parameters in wistar albino rats. International Journal of Biosciences, 2014; 4(7): 1-8.
- 23. Nafiu MO, Akanji MA, Yakubu MT; Effect of aqueous extract of Cochlospermum planchonii rhizome on some kidney and liver functional indices of albino rats. African Journal of Traditional Complementary and Alternative Medicine, 2011; 8(1): 22-26.
- 24. OCDE; Lignes directrices de l'OCDE pour les essais de produits chimiques: Étude de toxicité orale à dose répétée pendant 28 jours sur les rongeurs. OCDE 2008 ; 407 : 1-14.
- OMS; Stratégie de l'OMS pour la médecine traditionnelle pour 2000-2005. WHO/EDM/TRM, Génève; 2002: 1-65.
- 26. Pratt DS, Kaplan MM; Evaluation of abnormal liver-enzyme results in asymptomatic patients, New England Journal of Medicine, 2000; 342: 1266-1271
- Rosalki SB, Roberts R, Katus HA, Giannitsis E, Ladenson JH; Cardiac biomarkers for detection of myocardial infarction: Perspectives from past to present. Clinical Chimistry, 2004; 50: 2205-2213.
- Silbernagi S, Lang F; Atlas de poche de physiopathologie. 1<sup>ère</sup> édition, Flammarion Médecine-Sciences, Paris, 2000; 28-133.
- 29. Talke H, Schubert GE; Enzymatische harnstoffbestimmung in blut und serum im optischen test nach Warburb. Klinische Wochenschrift, 1965; 43: 174 -179.
- Tietz N; Fundamentals of Clinical Chemistry. Ed WB Saunders Co: Philadelphia 1987; 3: 373-401.
- Yelemou B, Bationo BA, Yaméogo G, Milogo-Rasolodimby J; Gestion traditionnelle et usage de *Piliostigma reticulatum* sur le plateau central du Burkina Faso. Bois et Forêts des Tropiques, 2007; 291(1): 55-66.