

Design and Development of Multiparticulate Floating Drug Delivery System Containing Lisinopril

Rathod Sayali P^{1*}, Jadhao Umesh T¹, Lokhande Sneha S¹, Panchal Pranita P¹, Wakade RB²¹Department of Pharmaceutics SDMVM's SVP College of Pharmacy Hatta, Parbhani, M.S Maharashtra, India²Department of Pharmaceutics SNIOP Pusad Dist-Yavatmal M.S Maharashtra, IndiaDOI: [10.36347/sajp.2021.v10i03.004](https://doi.org/10.36347/sajp.2021.v10i03.004)

| Received: 13.02.2021 | Accepted: 24.02.2021 | Published: 28.03.2021

*Corresponding author: Sayali P. Rathod

Abstract

Original Research Article

The aim of present work is to design multiple unit floating drug delivery system for prolong release of Lisinopril by using different amount of waxes, and to study effect of pectin on buoyancy of the system. The purpose of the study is to prepare wax-encorporated pectin-based emulsion gel beads using a modified Emulsion-gelation technique. The Model drug was incorporated in pectin wax contain olive oil, Lisinopril, were hot-melted, homogenized and then extruded into calcium chloride solution. The prepared Wax-incorporated Emulsion Gel Beads were evaluated for Micromeritics studies, entrapment efficacy, in-vitro buoyancy rate, dissolution rate, it was concluded that the increase of drug release of Lisinopril could be obtained upto 10 hrs. Various preformulation studies like bulk density, tapped density, Carr's index angle of repose were in the acceptable limits especially for batch F9. Drug and polymer are compatible with each other as they are verified through FTIR spectroscopy. Increasing the amount of wax in the formulation significantly prolonged the drug release but was insufficient for sustaining the release of highly water-soluble drug.

Keywords: Lisinopril, Multiparticulate, gastroretentive, buoyancy.

Copyright © 2021 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY-NC 4.0) which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

INTRODUCTION

Gastro retentive systems or dynamically controlled systems are low-density systems that have comparatively more buoyancy to float over the gastric contents and remain buoyant in the stomach without affecting the gastric emptying time for a prolonged period of rate & time. This results in an increased gastric retention time and a better control of the fluctuations in plasma drug concentration. Many buoyant systems have been developed based on granules, powders, capsules, tablets, laminated films and hallow Microspheres [1].

Gastro retentive drug delivery system (GRDDS) is one of the gastro retentive dosage forms which could prolong gastric retention time (GRT) to obtain sufficient drug bioavailability. This system floats in the gastric fluid due to its lower bulk density compared to that of the aqueous medium. FDSS is desirable for drugs with an absorption window in the stomach or in the upper small intestine. This system is also useful for drugs which act locally in the proximal part of gastrointestinal (GI) tract, such as antibiotic administration for *Helicobacter pylori* eradication in the

treatment of peptic ulcer and/or drugs which are poorly soluble or unstable in the intestinal fluid [2].

Multiparticulate systems show better reproducible pharmacokinetic behavior than conventional (monolithic) formulations. After disintegration which occurs within a few minutes often even subunits have diameters of less than 2 mm, they are able to leave the stomach continuously, even if the pylorus is closed. These results in lower intra and inter individual variability in plasma levels and bioavailability [3, 4].

It would, be advantageous to have means for providing an intimate contact of the drug delivery system with the absorbing membranes. This can be achieved by coupling gastroretentive and bioadhesion characteristics to multiparticulates and developing gastroretentive bioadhesive multiparticulates. These multiparticulates have advantages like efficient absorption and enhanced bioavailability of the drugs due to a high surface to volume ratio, a much more intimate contact with the mucus layer and specific targeting of drugs to the absorption site [5-8].

It is stated that, 'the multiparticulates' float on the stomach contents, and then adhere to the mucous linings as the stomach empties. The release of drug from the system can be controlled to coincide with the half-life emptying of the system from the stomach [9-11].

MATERIALS AND METHODS

Material

Lisinopril was obtained as kind gift sample from Yarrowchem Pharma, Mumbai India. Pectin purchased from Research lab fine chem. Industries, Mumbai, White wax & Olive oil was purchased from Thomas baker, Mumbai India. All other materials used of analytical grades.

Methods

Preparation of Conventional CaPG Beads and Emulsion Gel Beads

Conventional CaPG beads were prepared by the ionotropic gelation method 400 mg of pectin were dispersed in water with agitation, and then 100 mg of Lisinopril (80-mesh sieved) were dispersed in pectin solution to make a 100-g solution. The dispersion was then extruded through a plastic needle into 0.34 M calcium chloride which was gently stirred at room temperature. The gel beads formed were allowed to

stand in the solution for 20 min before being separated and washed with distilled water. The beads were dried at 37°C for 12 h.

The emulsion gel beads of calcium pectinate were prepared by emulsion-gelation method 400 mg of pectin were dissolved in water with agitation. 3 ml of olive oil were added to the mixture of pectin and lisinopril (1:1) to make 100-g mixtures and homogenized using a homogenizer, at 3,000 rpm for 5 min. The emulsion gel beads were treated in the same manner as conventional CaPG beads [12-13].

Preparation of Wax-incorporated Emulsion Gel Beads

Various amounts of different waxes (i.e. white wax and cetyl alcohol) were melted in water bath at 60–85°C, depending on the melting ranges of the waxes used. The molten wax was dispersed in the homogenized emulsion mixture of pectin, oil and Lisinopril which already heated to same temperature, and then mixed until the homogenous mixture was obtained. The hot melted mixture was extruded into 0.34 M calcium chloride (cooled at 5°C). The wax-incorporated emulsion gel beads obtained were treated in the same manner as conventional CaPG beads. The formulations studied are shown in Table

Table-1: Formulation Table

Batches	F1	F2	F3	F4	F5	F6	F7	F8	F9
Ingredients (mg)									
Lisinopril	100	100	100	100	100	100	100	100	100
Pectin	400	400	400	400	400	400	400	400	400
White wax	150	150	150	200	200	200	250	250	250
Cetyl alcohol	100	150	200	100	150	200	100	150	200
Olive oil (ml)	3	3	3	3	3	3	3	3	3
Distilled water	20	20	20	20	20	20	20	20	20
0.340M Cal. chloride (ml)	100	100	100	100	100	100	100	100	100

Evaluation of Lisinopril Beads

Micromeritics studies of floating beads [14]

The microspheres were characterized by their Micromeritics properties, such as particle size, bulk density, tapped density, Carr's compressibility index, hausner ratio and flow property.

$$\% \text{ Yield} = \frac{\text{Actual wt of product}}{\text{Total wt of expient and drug}} \times 100$$

Drug Entrapment Efficiency (DEE)

About 10g of beads were weighed accurately and crushed in glass mortar. Powdered beads were suspended in 100 ml of 0.1N HCL. After 24 hr the solution was filtered and filtrate was analysed for drug

$$\% \text{Drug entrapment} = \frac{\text{Calculated drug concentration}}{\text{Theoretical drug concentration}} \times 100$$

Percentage yield of microspheres formed.

The percentage yield of the prepared microsphere determined by weighing after drying. The measured weight of prepared microspheres was divided by the total amount of all the non-volatile components used for the preparation of the microspheres, which gave the total percentage yield of microspheres.

content at 230 nm. The results of % lisinopril loading and encapsulation efficiency were calculated using Equation. The drug content was calculated from standard curve.

In vitro buoyancy study: [16-18]

Appropriate quantity of the floating micro particulate was placed in 100 ml of the simulated gastric fluid (SGF, pH 2.0), the mixture was stirred with a magnetic stirrer. The layer of buoyant micro particulate was pipetted and separated by filtration. Particles in the sinking particulate layer were separated by filtration. Particles of both types were dried in a desiccator until constant weight was achieved. Both the fractions of microspheres were weighed and buoyancy was determined by the weight ratio of floating particles to the sum of floating and sinking particles.

$$\text{Buoyancy (\%)} = \frac{W_f}{W_f + W_s} \times 100$$

Where,

W_f and W_s are the weights of the floating and settled micro particles.

Dissolution test (in-vitro drug release) of microspheres

The dissolution test was performed using 900 ml 0.1N HCL fluid maintained at $37 \pm 0.50^\circ\text{C}$ and stirred at 100 rpm for 10 hr. A sample (1 ml) of the aliquots were withdrawn at different time intervals and an equivalent volume of medium prewarmed at 37°C was added to maintain sink condition. The withdrawn samples were filtered through Whatman filter paper no. 41. Absorbance of these solutions was measured at 230 nm. Percentage drug release was calculated using an equation obtained from a standard curve.

Infrared Spectroscopy Interpretation for interaction between drug and polymer in Microsphere

Fourier transforms infrared spectroscopy (FTIR) spectra of the pure drug and the microspheres were produced using by KBr disk method. Powder microsphere and the ingredients used in drug loading were subjected to FTIR with a Shimadzu 8201 PC FTIR. Background spectrum was collected before

running each sample. The samples were analyzed between wave numbers 4000 and 400 cm^{-1} [20-22].

Stability Study for Multiple Unit System

During the stability studies the product is exposed to normal conditions of temperature and humidity. However the studies will take a longer time and hence it would be convenient to carry out the accelerated stability studies where the product is stored under extreme conditions of temperature. In the present study, stability studies were carried out on optimized formulation. The microspheres were stored at $40 \pm 20^\circ\text{C}/75 \pm 5\% \text{ RH}$ for duration of one month. At the interval after thirty days each sample was withdrawn and tested for drug entrapment, and drug release study.

RESULT AND DISCUSSION**Micromeritic Properties**

Micromeritic properties for batch F1 to F9 are shown in Table No. 9. The results showed bulk density and Carr's index in the range of 12% to 16%, Hausner's ratio less than 1.25 which shows good flow properties. The values were found to be in the range of $21^\circ 28' \pm 1.4$ to $29^\circ 69' \pm 1.8$, beads shows the angle of repose less than 30° which reveals good flow property. The observed results suggest good flow ability of the beads. Bulk density may affect buoyancy of floating beads. The bulk density of formulations was found to be between 0.165 ± 0.02 to $0.254 \pm 0.02\text{ g/cm}^3$. This indicates good packing capacity. The tapped density of beads was found to be in range of 0.197 ± 0.032 to $0.308 \pm 0.018\text{ g/cm}^3$, which shows good packability of beads. The mean particle sizes of all formulations were ranged from $2.04 \pm 0.37\text{ mm}$ to $2.32 \pm 0.60\text{ mm}$ as shown in Table No 10. Higher particle size was obtained when the proportion of waxes was increased in polymer and drug mixture of pectin: lisinopril. As the amount of calcium chloride was increased, more crosslinking structure was observed that lead to a decrease in particle size.

Table-2: Micromeritics studies of floating beads

Para-meters Batches	Bulk density (g/cm^3)	Tapped density (g/cm^3)	Carr's Index	Hausner ratio	Angle of repose (θ)	Particle size (mm)
F1	0.246 ± 0.02	0.308 ± 0.018	20.12 ± 0.15	1.25 ± 0.12	$28^\circ 65' \pm 0.8$	2.23 ± 0.35
F2	0.165 ± 0.02	0.197 ± 0.032	16.24 ± 0.12	1.19 ± 0.006	$26^\circ 50' \pm 2.5$	2.2 ± 0.43
F3	0.186 ± 0.02	0.219 ± 0.041	15.06 ± 0.26	1.17 ± 0.005	$29^\circ 69' \pm 1.8$	2.24 ± 0.35
F4	0.236 ± 0.03	0.265 ± 0.027	10.94 ± 0.18	1.12 ± 0.006	$27^\circ 23' \pm 1.5$	2.22 ± 0.19
F5	0.222 ± 0.01	0.255 ± 0.019	12.94 ± 0.20	1.14 ± 0.007	$28^\circ 65' \pm 0.8$	2.04 ± 0.37
F6	0.254 ± 0.02	0.306 ± 0.029	16.99 ± 0.23	1.20 ± 0.004	$26^\circ 50' \pm 2.1$	2.15 ± 0.222
F7	0.250 ± 0.03	0.283 ± 0.04	11.66 ± 0.21	1.13 ± 0.002	$26^\circ 69' \pm 2.3$	2.32 ± 0.60
F8	0.217 ± 0.01	0.261 ± 0.030	16.85 ± 0.24	1.20 ± 0.004	$21^\circ 28' \pm 1.4$	2.26 ± 0.23
F9	0.225 ± 0.02	0.257 ± 0.023	12.45 ± 0.19	1.14 ± 0.007	$27^\circ 23' \pm 1.2$	2.26 ± 0.20

Determination of Percentage yield

It was found that the average percentage yield was greater than 80% for all formulations. The drug loading was found to be in range of $84.21 \pm 0.5\%$ to

$97.22 \pm 0.2\%$. Formulation F1 showed highest loading of $97.22 \pm 0.2\%$. Whereas formulation F9 showed lowest drug loading of $84.21 \pm 0.5\%$. Overall the drug loading

was decreased with increase in the concentration of waxes.

Drug entrapment efficiency

Ionic gelation technique is convenient method for the preparation of floating beads with good drug loading and encapsulation efficiency. In this method drug is dispersed equally in the polymer matrix so drug can be loaded easily in the polymer. The drug entrapment efficiency of the prepared beads was found to be increased progressively with an increase in

concentration of waxes and polymers. This might be due to increased matrix density.

In vitro buoyancy study

The formulation F1 gives best floating ability of 92.38% in 0.1 N HCL for 10 hrs. This may be due to its low bulk density. The floating ability of the formulations ranges from 65.52% to 93.20%, which shows the excellent floating ability of the beads. In this as the concentration of wax increases, the viscosity of mixture increases and thus leads to good floating ability.

Table-3: Percentage yield, In vitro buoyancy study, Drug entrapment efficiency

Batch	Drug entrapment efficiency (%)	Yield (%)	(%) buoyancy
F1	86.45%	97.22±0.2	92.38%
F2	93.20%	94.11±0.7	84.34%
F3	88.67%	92.94±0.2	90.67%
F4	76.25%	95±0.4	79.33%
F5	65.52%	86.87±0.5	69.32%
F6	83.34%	91.73±0.3	78.34%
F7	75.38%	96.47±0.3	86.12%
F8	76.50%	88.88±0.4	83.78%
F9	89.42%	84.21±0.5	77.23%

In vitro drug release study

It was observed that the beads ascended to the upper part of the dissolution vessels and remains floated until the completion of release studies. The drug release study was carried for 10 hrs. This showed as the concentration of waxes increases the release rate of the drug decreases which were attributed to increase density of the polymer matrix at highest concentration causing decreased diffusional path length, and increases

drug release from the polymer matrix. Moreover dissolution study data revealed that release from the beads was largely dependent on the polymer swelling and drug diffusion. The percentage drug release from batches F1 to F9 varied from 61.01 % to 96.31 %. The in-vitro drug release of the formulation F1 to F9 displayed in Table where comparative release of drug is shown.

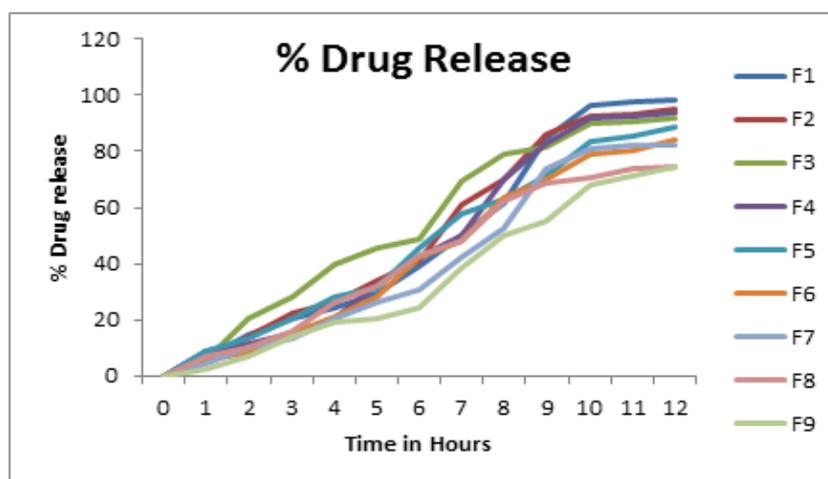


Fig-1: In vitro drug release study

IR Study

The infrared absorption spectrum of pure lisinopril and the sustained release microsphere of optimized polymer were recorded on FT-IR spectrophotometer (Model-8400S, Shimadzu, Japan) and the spectrum analysis was done for functional groups. The FT-IR spectra of drug with polymer were

compared with the standard FT-IR spectrum of the pure drug. 1658, 1611 (the vibrations characteristic to carboxyl group), 1578 cm^{-1} (NH bending group vibration) and 1546 cm^{-1} for pure lisinopril compound are shifted to 1637 and 1592 cm^{-1} for all IC products. These frequency shifts can be explained probably by breaking the hydrogen bonds in the case of NH groups

and respectively by the formation of hydrogen bonds in the case of car-boxyl groups. Consequently, FTIR studies established clearly the functional groups implied in the inclusion process. In the 4000–2500 cm⁻¹ spectral region, see Fig. 4, the most important

contributions are due to the O–H stretching vibrations of primary (3504 cm⁻¹[9]), secondary OH groups of the groups and soft water molecules present inside CD cavity, also. The different shape of this mass if (corresponding to ICs’).

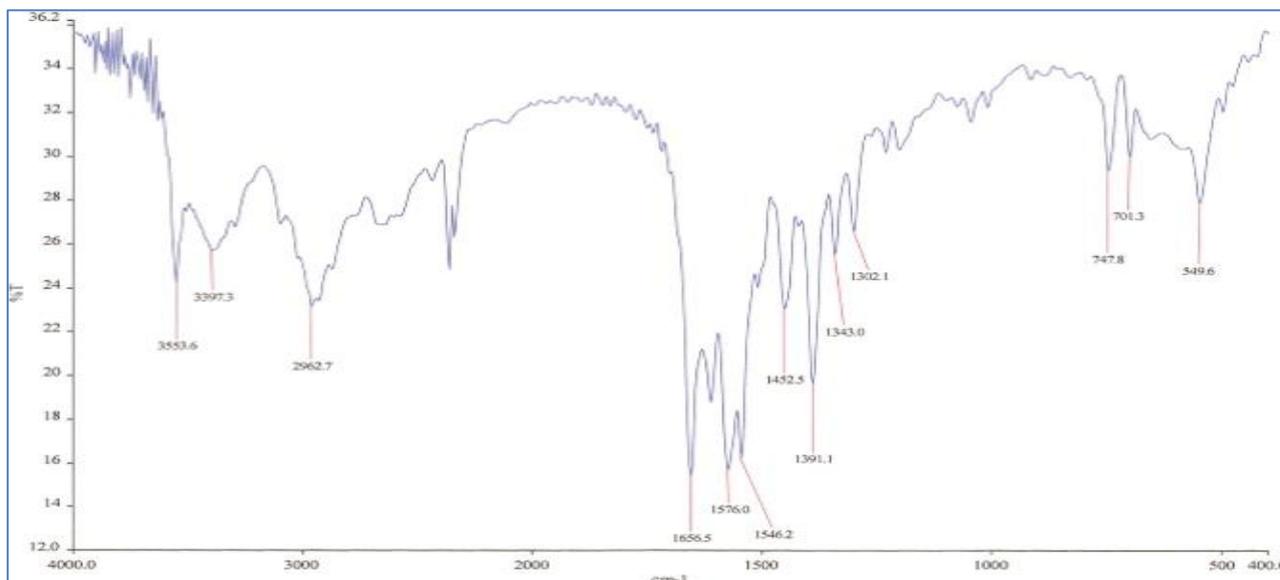


Fig-2: FTIR of Lisinopril

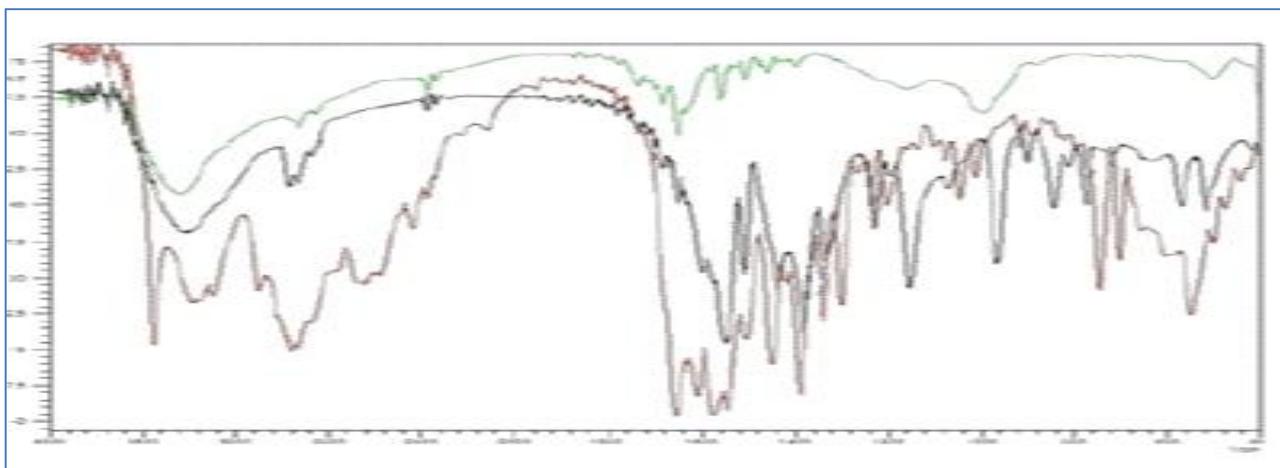


Fig-3: FTIR of Lisinopril physical mixture

CONCLUSION

Intragastric floating drug delivery system was prepared by incorporating low density materials, e.g. oils and/or waxes. Incorporation of wax into the beads influenced the drug release. It can be concluded that the drug Lisinopril multiparticulate floating beads for the treatment of hypertension were prepared and the release kinetics of the optimized batch could show the assured sustain release of drug. By using waxes it was concluded that the increase of drug release of Lisinopril could be obtained upto 10 hrs. Various preformulation studies like bulk density, tapped density, Carr's index angle of repose were in the acceptable limits especially for batch F9. Drug and polymer are compatible with each other as they are verified through FTIR

spectroscopy. Increasing the amount of wax in the formulation significantly prolonged the drug release but was insufficient for sustaining the release of highly water-soluble drug.

REFERENCES

1. Arora S, Ali J, Khar RK, Baboota S, Floating drug delivery systems: A review, *AAPS Pharm Sci Tech.* 2005;6(3): 372-390.
2. Babu VBM, Khar RK, In vitro and In vivo studies of sustained release floating dosage forms containing salbutamol sulphate, *Pharmazie.* 1990; 45: 268-270.
3. Bardonnnet PL, Faivre V, Pugh WJ, Piffaretti JC and Falson F, Gastro retentive Dosage Forms:

- Overview and Special case of *Helicobacter pylori*. Journal of Control Release. 2006; 111.
4. Bari PH. A Comprehensive review on gastro retentive drug delivery system, IPP. 5(2):94-102,2017
 5. Chandiran S, Kumar BP and Narayan V., Formulation and in vitro evaluation of floating drug delivery system for salbutamol sulphate, International Journal of Pharma Biomed Sciences. 2010; 1(1): 12-15.
 6. Chein YW, "Novel Drug Delivery System" 2nd ed. Marcel decker Inc., New York, 1-3.
 7. Cook JD, Carriaga M, Kahn SG, Schalch W, Skikne BS, Gastric delivery system for iron supplementation, Lancet. 1990; 335: 1136-1139.
 8. Abdul Sayeed, Gastro retentive Drug Delivery Systems: A Review, Der Pharmacia Lettre. 2011, 3(1): 121-137.
 9. Streubel A, Siepmann J, Bodmeier R. "Floating microparticles based on low density foam powder", Int J Pharm. 2002; 241:279-92.
 10. Desai S, Bolton S, A floating controlled release drug delivery system: in vitro- in vivo evaluation. Pharm Res. 1993, 10(9): 1321-1325.
 11. Desai S. A Novel Floating Controlled Release Drug Delivery System Based on a Dried Gel Matrix Network [master's thesis]. Jamaica, NY: St John's University; 1984.
 12. Dixit N, Floating Drug Delivery -System, Journal of Current Pharmaceutical Research. 2011; (1): 6-20.
 13. Tekade BW. Optimization and in-vitro evaluation of verapamil hydrochloride floating tablet. The pharma innovation. 2014,3(6): 42-48
 14. Garg S and Sharma S, Gastroretentive Drug Delivery System, Business Briefing: Pharma tech. 2003; 7:160-166.
 15. Tekade BW, Jadhao UT, Thakare VM, Bhortake LR. Formulation and evaluation of diclofenac sodium effervescent tablet. Infrared Spectroscopy. 2014;9(10):11.
 16. Geetha A, Kumar J Rajendra and Mohan CH Krishna, Review on: Floating drug delivery systems, International journal of pharmaceutical research and biomedical analysis. 2012; (1): 1-13.
 17. Grubel P, Gastric emptying of non-digestible solids in the fasted dog, J Pharm Sci., 1987;76: 117 – 122.
 18. Jadhao UT, Effect of Excipients and Process Variables over Gastro Retentive Antihypertensive Dosage Form, International Journal of Pharmaceutical Research & Analysis. 2014; (4)3:186-192.
 19. Jadhao UT. Design and Evaluation of Famotidine Matrix Tablet Using 3² Factorial Designs, RJPBCS. 2013;3(4),1441-1451.
 20. Jadhav CM, Bendale AN, Patil VR, Tekade B W, Thakare VM, Formulation and Evaluation of Effervescent Floating Tablet of Felodipine, IJPRD. 2012; 3(12): 43-48.
 21. Jain A. New Concept: Floating Drug Delivery System, Indian Journal of Novel Drug Delivery. 2011; 3(3): 163-69.
 22. Watson DG. Pharmaceutical Analysis A textbook for pharmacy students and pharmaceutical chemists, first ed. London, Churchill Livingstone. 1999: 100-03.
 23. Duerst M. Spectroscopic methods of analysis: infrared spectroscopy. Swarbrick J., Boylon JC, Encyclopedia of Pharmaceutical Technology. 2007; 3:3405-18.
 24. Skoog DA, Holler FJ, Nieman TA. Principles of Instrumental Analysis. 5 th ed. Sounder's College Publishing. 2004: 798- 808.