

## Clinico Microbiological Spectrum of Diabetic Foot Infections

Dr. Mohamed Jan<sup>1</sup>, Dr. Haja Abdul Nazeer MJ<sup>2\*</sup>

<sup>1</sup>Assistant professor, Dept of Gen Surgery, Vinayaka missions medical college and hospital, Karaikal, Puducherry, India

<sup>2</sup>Professor, Dept of Microbiology, Vinayaka missions medical college, Karaikal, Puducherry, India

### Original Research Article

\*Corresponding author

Dr. Haja Abdul Nazeer MJ

#### Article History

Received: 01.03.2018

Accepted: 07.03.2018

Published: 30.03.2018

#### DOI:

10.36347/sjams.2018.v06i03.076



**Abstract:** Diabetic foot infections are a common cause of morbidity and mortality. This study was undertaken to study the microbiological flora of diabetic foot ulcers and to assess the in vitro antibacterial susceptibility of the causative bacteria. This was a prospective observational study conducted in which 247 wound swabs collected from patients with diabetic foot infection. All collected specimens processed using standard microbiological techniques. Disc diffusion method was used to find out the susceptibility of bacterial agents. 209 specimens yielded microbial growth on culture media. Monomicrobial growth accounted for 74.06%. Staphylococcus aureus was the predominant organism isolated in which MRSA was 27%. Second common pathogen was Pseudomonas aeruginosa. All Gram positive strains were found sensitive to linezolid. Imipenem showed good susceptibility against Gram negative bacilli. Candida was the only fungi isolated. Staphylococcus aureus was isolated predominantly from diabetic foot infections. Monomicrobial growth was seen in 177 patients and accounted for 74.06%. Polymicrobial growth was seen in 62 patients and accounted for 25.94%. The most common combination was S. aureus and Candida species. Majority of Gram positive and Gram negative isolates were susceptible to linezolid and imipenem respectively.

**Keywords:** Diabetic foot infections, Staphylococcus aureus, Linezolid.

### INTRODUCTION

Diabetes mellitus is a metabolic disorder characterized by chronic hyperglycemia and target organ damage.

Diabetes is a fairly common disease seen in India with a prevalence of almost 12% - 17% in the Indian urban population as per a study in 2001 with a prevalence of 2.5% in the rural population [1]. The Indian diabetic population is expected to increase to 57 million by the year 2025 [2]. Various Co morbidities with diabetes mellitus are cardiovascular disease, retinopathy, nephropathy, neurological, peripheral vascular diseases and infections. Approximately 15% of patients with DM develop foot infections which eventually progress to osteomyelitis and amputation [3].

A commonly accepted definition of foot infection is the presence of systemic signs of infection (e.g., fever and leukocytosis) or purulent secretions or two or more local symptoms or signs (redness, warmth, indurations, pain, or tenderness) [4]. Different studies have reported on microbiological spectrum of Diabetic Foot Infections (DFIs) over the past 25 years, but the results have been varied and often contradictory [5]. S.aureus, Enterococci, E.coli, Proteus and Pseudomonas spp. are the most frequent pathogens which are cultured from diabetic foot ulcers. However, the

etiology of wound infection differs from country to country and from hospital to hospital even within the same region [6]. Many of these microorganisms are developing resistance to commonly used antibiotics largely due to their inappropriate use.

Hence, this study was undertaken to study the microbiological flora of diabetic foot ulcers and to assess the in vitro antibacterial susceptibility of the causative bacteria.

### MATERIALS AND METHODS

This was a prospective observational study in which 247 consecutive samples from diabetic patients with foot ulcers attending outpatient and in patient department of general surgery of vinayaka mission's medical college and hospital over a period of six months i.e from July 2017 to December 2017 were included, after getting the informed consent from the study group. All the patients underwent detailed history and clinical examination. Demographical data that included age, sex, duration of diabetes, duration of

diabetic foot, location of foot ulcer, and Wagner’s grade were recorded for every case.

**Sample collection**

Wound beds were prepared before specimen collection, where the wound immediate surface exudates and contaminants were cleansed off with moistened sterile gauze and sterile normal saline solution. Dressed wounds were cleansed with non-bacteriostatic sterile normal saline after removing the dressing. Aseptically the end of a sterile cotton-tipped applicator was rotated over 1 cm<sup>2</sup> area for 5 seconds with sufficient pressure to express fluid and bacteria to surface from within the wound tissue. Two wound swabs were taken from each wound at a point in time to reduce the chance of occurrence of false-negative cultures and to increase the chance of recovering bacterial pathogens. It is also indicative of contamination in that if the two swab samples differ in types of organisms during presumptive test [7].

The specimens were subjected to gram staining 10% KOH and they were inoculated onto

bacteriological medium (Blood agar, Mac Conkey’s agar, chocolate agar) and mycological medium (SDA) for the isolation of aerobic bacteria and fungi respectively. The isolates were identified by the standard biochemical tests. Antibiotic sensitivity testing was performed by Kirby Bauer-Disk Diffusion Method [8].

Inclusion/Exclusion criteria: Patients who had received systemic antibiotic therapy for more than 24 h within the previous 72 h and those who had undergone amputation were excluded from the study.

**RESULTS**

A total of 247 patients who had diabetic foot infections were enrolled in the study. 209 specimens yielded microbial growth on culture media. Monomicrobial growth was seen in 177 patients and accounted for 74.06%. Polymicrobial growth was seen in 62 patients and accounted for 25.94%. Demographic details of patients shown in Table-1.

**Table-1: Demographic details of patients with diabetic foot infections**

Demographic data	NUMBER (%)
Age	
Sex	
Male	140 (66.99%)
Female	69 (33.01%)
Duration of diabetes	
< 1 year	50 (23.92%)
> 1 year	159 (76.08%)
Duration of Ulcer	
< 1 month	119 (56.94%)
>1 month	90( 43.06%)
Grading of ulcer Wagner’s classification	
Grade 2	57 (27.27%)
Grade 3	78 (37.32%)
Grade 4	43 (20.57%)
Grade 5	31 (14.83%)

Staphylococcus aureus (32.22%) was the predominant organism isolated from diabetic foot. Pseudomonas aeruginosa (24.68%) was the second common pathogen isolated followed by Enterococci

(12.55%) and Klebsiella species (12.13%). In the present study, Candida species (6.69%) was the only fungi isolated from diabetic foot infections (Table-2)

**Table-2: Organisms isolated from diabetic foot**

Organism	Number (%)
Stapylococcus aureus	77 (32.22%)
Pseudomonas aeruginosa	59 (24.68%)
Enterococci	30 (12.55%)
Klebsiella species	29 (12.13%)
E.coli	21 (8.79%)
Proteus species	5 (2.09%)
CONS	2 (0.84%)
Candida species	16 (6.69%)
Total	239(100%)

CONS: Coagulase negative staphylococci.

In our study, Methicillin resistant Staphylococcus aureus (MRSA) was isolated and accounted for 27%. All MRSA strains were susceptible to linezolid (100%) followed by

piperacillin/tazobactam (95%). Ceftazidime (9%) and Ceftriaxone (4%) were found to least susceptible. All Gram positive cocci were found to be susceptible to linezolid (100%) (Table-3).

**Table-3: Susceptibility pattern of Gram positive cocci**

Antibiotic	MRSA(n=21)	MSSA(n=46)	Enterococci(n=30)	CONS(n=2)
Ampicillin	0(0)	9(19%)	10(33%)	0(0)
Amikacin	11(52%)	41(89%)	NT	2(100%)
Gentamicin	5(23%)	21(45%)	NT	2(100%)
Ciprofloxacin	5(23%)	30(65%)	8(26%)	1(50%)
Pipracillin/Tazobac	20(95%)	44(95%)	17(56%)	2(100%)
Cefeperazone/Sulbact	11(52%)	35(76%)	15(50%)	2(100%)
Ceftazidime	2(9%)	18(39%)	11(36%)	1(50%)
Ceftriazone	1(4%)	17(36%)	10(33%)	1(50%)
Linezolid	21(100%)	46(100%)	30(100%)	2(100%)

MRSA: Methicillin resistant Staphylococcus aureus

MSSA: Methicillin sensitive Staphylococcus aureus

CONS: Coagulase negative Staphylococci.

NT: Not tested

Imipenem was most susceptible antibiotic to majority of Gram negative strains, except Pseudomonas aeruginosa (94%) and E.coli (95%).

Piperacillin/tazobactam and Cefeperazone/sulbactam showed good susceptibility (93%-100%).

**Table-3: Susceptibility pattern of Gram negative bacilli ()**

Antibiotic	Pseudomonas(n=59)	Klebsiella (n=29)	E.coli(n=21)	Proteus(n=5)
Amikacin	50(84%)	28(96%)	21(100%)	5(100%)
Gentamicin	25(42%)	11(37%)	17(80%)	5(100%)
Ciprofloxacin	21(35%)	19(65%)	15(71%)	3(60%)
Pipracillin/Tazobac	55(93%)	28(96%)	20(95%)	5(100%)
Cefeperazone/Sulbact	51(86%)	28(96%)	20(95%)	5(100%)
Ceftazidime	31(52%)	12(41%)	9(42%)	4(80%)
Ceftriazone	19(32%)	14(48%)	3(14%)	2(40%)
Imipenam	56(94%)	29(100%)	20(95%)	5(100%)

**DISCUSSION**

Diabetic foot pathologies are common in diabetics and pose serious health problems for developing countries. In the present study, males (66.99%) were predominantly developed diabetic foot infections. Similar findings were observed in other studies that male sex has been purported to be a risk factor for the development of diabetic foot lesions [9]. But there is one study by Bose [10] reporting female patients to constitute the majority of the patients. The mean age of patients in the present study is 55.35 + 17.5. While the study of Benedicto *et al.*, showed an incidence of diabetic foot ulcer with mean age 68 + 5.9 [11].

In the present study, diabetic foot infections were predominantly developed among diabetic patients who had diabetes more than one year (76.08%). In our study, majority of diabetic patients who developed foot infections were grade 3(37.32%). According to the study conducted by Priyadarshini *et al.*, maximum

number of patients with infected diabetic foot ulcers belonged to Wagner grade 3 and 4 [12]. Monomicrobial growth was observed in 74% patients. But as per Priyadarshini *et al.*, [12] monomicrobial growth was seen in 50% patients. Mohd Zubair *et al.*, [13] Anandi *et al.*, [14] Rama Kant *et al.*, [15] Pappu K *et al.*, [16] and Citron *et al.*, [5] have reported 56.6%, 19%, 23 %, 92% and 16.2 % monomicrobial infections and 33%, 67%, 66%, 7.7% and 83 % of polymicrobial infections respectively. The most common combination was found to be Staphylococcus aureus and Candida species. According to the study conducted by Saravanan Sanniyasi *et al.*, [17] commonest bacterial organism found in fungal positive patients was Pseudomonas followed by Enterococcus compared to the study by Emilija *et al.*, [18] where Enterobacter followed by Pseudomonas were the commonest organisms.

In our study, S.aureus (32%) was the predominant pathogen isolated. This is similar to the studies conducted by Citron *et al.*, [5] Mohammed

Zubair *et al.*, [13], and Alavi SM *et al.*, [19] reported *Staphylococcus aureus* as the predominant pathogen, which comprised 57.2%, 28% and 26.2% of their isolates respectively. In our study, Enterococci was accounted for 12%. But as per Citron *et al.*, [5] and Mohammed Zubair *et al.*, [13] *Streptococcus pyogenes* was isolated in 10% and 15% respectively. In contrast, Pappu K *et al.*, [16], who reported that 76% of the organisms which were isolated were gram negative bacilli, *Pseudomonas* being the predominant pathogen (23%), followed by *Staphylococcus aureus* (21%). Mohammed Zubair *et al.*, [13] reported *Escherichia coli* (26.6%) and *Pseudomonas aeruginosa* (10.6 %) as the predominant gram negative isolates.

In the present study, 16 specimens from diabetic foot infections yielded the growth of *Candida* species and accounted for 6% which is lower compared to Bansal E *et al* (20) 9% (9 out of 103 patients) and higher compared to Emilija *et al.*, [18] 4.5% (23 out of 509 patients). Emilija *et al.*, [18] grew only *Candida* species which is similar to the present study. But other studies reported *Aspergillus* (30%) [21].

In our study, no anaerobic bacterial culture was performed. Involvement of anaerobic bacteria in diabetic foot infections is not clear and few studies reported minor role of anaerobic bacteria [22]. While other studies reported preponderance of anaerobic bacteria [23].

In this study, 27% MRSA were isolated. But Umadevi *et al.*, [24] reported 65% MRSA from diabetic foot infections. While other studies on diabetic foot infections which have reported 10–44% MRSA [9]. MRSA are more often isolated from patients who have been previously hospitalized or reside in a chronic care facility, who have recently received antibiotic therapy or who have had a previous amputation. The isolation of MRSA in DFIs would be associated with more severe infections [24].

## CONCLUSION

*Staphylococcus aureus* was isolated predominantly from diabetic foot infections. Monomicrobial growth was seen in 177 patients and accounted for 74.06%. Polymicrobial growth was seen in 62 patients and accounted for 25.94%. The most common combination was *S.aureus* and *Candida* species. Linezolid was found to be sensitive to all Gram positive cocci including MRSA. Majority of Gram positive and Gram negative isolates susceptible to Piperacillin /tazobactam and amikacin.

## REFERENCES

1. Vasista SG. Epidemiology of diabetes mellitus urban-rural –A paradox. In: Jayaram BM, editor. Type II Diabetes: Urban–Rural. 1st ed. Bangalore : Microlabs Ltd; 2004.p 24-25.
2. Shakil S, Khan AU. Infected foot ulcers in male and female diabetic patients: A clinico-bioinformative study. *Ann Clin Microbiol Antimicrob.* 2010; 9:1-10.
3. Ramsey SD, Newton K, Blough D, McCulloch DK, Sandhu N, Reiber GE, et al. Incidence, outcomes, and cost of foot ulcers in patients with diabetes. *Diabetes Care* 1999;22:287–382.
4. Kahn, R. “Consensus development conference on diabetic foot wound care.” *Diabetes Care*, vol. 22, no. 8, pp. 1354–1360, 1999.
5. Citron DM, Goldstein EJC, Merriam VC, Lipsky BA. Bacteriology of moderate to severe diabetic foot infections and invitro activity of antimicrobial agents. *J Clin Microbiol.* 2007; 45 (9):2819–28.
6. Biadlegna F, Abera B, Alem A, Anagaw B. Bacterial isolates from wound infection and their antimicrobial susceptibility pattern in Felege Hiwot referral hospital North West Ethiopia. *Ethiop J Health Sci* 2009;19:173-9.
7. Bori G, Soriano A, Garcia S, Mallofre C, Riba J, Mensa J. Usefulness of histological analysis for predicting the presence of microorganisms at the time of reimplantation after hip resection arthroplasty for the treatment of infection. *J Bone Joint Surg Am* 2007;89:1232-61.
8. Forbes BA, Sahm DF, Weissfeld AS. Chapter 13, Overview of bacterial identification methods and strategies. *Bailey and Scott’s diagnostic Microbiology*, 12th ed. St. Louis: Mosby; 2007. p. 216-47.
9. Gadepalli R, Dhawan B, Sreenivas V, Kapil A, Ammini AC, Chaudhry R. A clinico-microbiological study of diabetic foot ulcers in an Indian tertiary care hospital. *Diabetes care.* 2006 Aug 1;29(8):1727-32.
10. Bose K. Infection in diabetic foot. *Ann Acad Med Singapore* 1978, 7: 359–365.
11. Benedicto J, Juarez R, Lim S. A descriptive study on the clinical characteristics, management and the outcome of diabetic patients with foot lesions admitted at the Philippine General Hospital. *Phil J Intern Med.* 1996;34:185–192.
12. Shanmugam P, Jeya M. The bacteriology of diabetic foot ulcers, with a special reference to multidrug resistant strains. *Journal of clinical and diagnostic research: JCDR.* 2013 Mar;7(3):441.
13. Zubair M, Malik A, Ahmad J. Clinico-bacteriology and risk factors for the diabetic foot infection with multidrug resistant microorganisms in North India. *Biol Med.* 2010; 2 (4): 22-34.
14. Anandi C, Alaguraja D, Natarajan V. Bacteriology of diabetic foot lesions. *Indian J Med Microbiol.* 2004; 22 (3): 175 – 78.
15. Ramakant P, Verma AK, Misra R, Prasad KN. Changing Microbiological profile of pathogenic bacteria in diabetic foot infections: time to rethink on which empirical therapy to chose? *Diabetologica.* 2011; 54 (1): 58-64.

16. Pappu AK, Sinha A, Johnson A. Microbiological profile of diabetic foot ulcer. *Calicut Med Journal*. 2011; 9(3):e:1-4.
17. Sanniyasi S, Balu J, Narayanan CD. Fungal Infection: A Hidden Enemy in Diabetic Foot Ulcers. *J Foot Ankle Surg (Asia-Pacific)* 2015;2(2):74-76.
18. Mlinaric-Missoni E, Kaenic S, Vukelic M. Candida infections of diabetic foot ulcers. *Diabetologia Croatica* 2005;34(1):29-35.
19. Alavi SM, Khosravi AD, Sarami A, Dashtbozorg A, Montazeri EA. Bacteriologic study of diabetic foot ulcer. *Pak J Med Sciences*. 2007;23(5):681–84.
20. Bansal E, Garg A, Bhatia S, Attri AK, Chander J. Spectrum of microbial flora in diabetic foot ulcers. *Ind J Pathol Microbiol* 2008;51(2):204-208
21. Raza M, Anurshetru BS. Clinical study of coexistence of fungal infections in diabetic foot ulcers and its management. *Int Surg J* 2017;4:3943-50.
22. Senneville E, Melliez H, Beltrand E, Legout L, Valette M, Cazaubiel M. Culture of percutaneous bone biopsy specimens for diagnosis of diabetic foot osteomyelitis: concordance with ulcer swab cultures. *Clin Infect Dis* 2006;42:57-62.
23. Abdulrazak A, Bitar ZI, Al-Shamali AA, Mobasher LA. Bacteriological study of diabetic foot infections. *J Diab Comp* 2005;19:138-141.
24. Umadevi S, Kumar S, Joseph NM, Easow JM, Kandhakumari G, Srirangaraj S, Raj S, Stephen S. Microbiological study of diabetic foot infections. *Indian Journal of Medical Specialities* 2011;2(1):12-17.
25. Tentolouris N, Petrikkos G, Vallianou N, Zachos C, Daikos GL, Tsapogas P, Markou G, Katsilambros N. Prevalence of methicillin-resistant *Staphylococcus aureus* in infected and uninfected diabetic foot ulcers. *Clinical microbiology and infection*. 2006 Feb 1;12(2):186-9.