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Ophthalmology

# Aspergillus niger as a Cause of Conjunctival Lithiasis in Children

Dr. Kavitha Toopalli<sup>1\*</sup>, Dr. Modini Pandharpurkar<sup>2</sup>

<sup>1</sup>Associate Professor of Pathology, Gandhi Medical College, Hyderabad, Telangana, India <sup>2</sup>Professor of Ophthalmology, Sarojini Devi Eye Hospital, Hyderabad, India

	Abstract. Conjunctival lithiasis in children is very rare, with a single case reported
	in literature to the best of our knowledge. The present study constituted twelve
<u>Uriginal Research Article</u>	In interature to the best of our knowledge. The present study constituted twerve
	cases of children presenting to Sarojini Devi Eye Hospital, Hyderabad between
*Corresponding author	December 2013 to November 2017 with the complaint of formation of stones in
Dr. Kavitha Toopalli	their eyes. The cytological examination of material formed in the lower
Dr. Havina 100pani	conjunctival fornix and that of the crushed stone was done. Microbiological
Article History	culture and biochemical analysis of the stones was done. Conjunctival biopsies
Received: 30.08.2018	taken from the lower conjunctival fornix were fixed in 10% formalin, processed
Accepted: 10.09.2018	routinely, stained with Hematoxylin & Eosin and special stains. Stones were sent
Published: 30.09.2018	for biochemical analysis. The cytology, histopathology, and microbiology and
1 ubusneu. 50.07.2010	biochemical findings are correlated. The cytohistological findings revealed
DOI	branched fungal hyphae, calcium oxalate crystals and dark brown to black pigment
10 36347/siams 2018 v06i09 034	deposits. A diagnosis of Aspergillusniger was made. Microbiological culture
10.50547/Sjams.2010.00007.054	showed fungal growth consistent with Aspergillusniger on microscopy.
real-user of real	Biochemical analysis showed that the stones were composed of calcium
「国家議会」「国	compounds. Aspergillus niger releases a mycotoxin, oxalic acid which complexes
2.7 5 36	with calcium ions in tear fluid to precipitate as calcium oxalate crystals.
	Crystallogenesis is essential to stone formation. The probable mechanism of
1528320	conjunctival stone formation and possible factors responsible for the rapid
TET 2049446	formation of conjunctival stones are analysed
	formation of conjunctival stones are analysed.

**Keywords**: Conjunctival lithiasis, children, *Aspergillusniger*, oxalic acid, calcium oxalate crystals.

# INTRODUCTION

Aspergillus species is the most ubiquitous fungi seen in soil, water & decaying vegetations.<sup>[1]</sup> Most infections are attributed to *Aspergillusfumigatus*, *flavus* and *terreus*.<sup>[2]</sup> They affect the lungs, naso-orbital area, skin and may be disseminated [1]. *A. niger* is an uncommon cause of invasive aspergillosis. Concretions occurring as minute hard yellow spots in the palpebral conjunctiva due to accumulation of epithelial cells and inspissated mucus in depressions called Henle glands is sometimes seen in elderly people who suffered from trachoma.However rapid and spontaneous formation of stones of varying sizes in the conjunctiva of children is quite rare and puzzling. This study is taken up to know the underlying cause of conjunctival stone formation in children that helps the clinician plan specific treatment.

### **MATERIALS & METHODS**

The present study constituted twelve cases of children presenting to Sarojini Devi Eye Hospital, Hyderabad between December 2013 to October 2017 with the complaint of formation of stones in their eyes. All the children came from lower socio-economic strata. The number of conjunctival stones formed on an average ranged from one to ten per day. The stones formed intermittently for few days with no stone formation for few days. There was no relevant past or family history. Routine hematological and biochemical investigations were done on all the patients. Soft material that appeared in the lower conjunctival fornix prior to hardening was smeared on slides and fixed in 90% alcohol. The hard stones were cleaned in xylene, crushed and made into fine powder, resuspended with egg albumin and smeared on slides. Cytology smears were stained with hematoxylin & eosin and Papanicoulou stains. Microbiological culture of the material and stones was done. Few stones were sent for biochemical analysis. Conjunctival biopsies from the lower fornix were submitted to histopathology. Formalin fixed tissues were routinely processed. Sections of four microns were cut from the paraffin wax embedded tissue blocks and stained with Hematoxylin & Eosin. Special stains, Gomori's Methanamine Silver (GMS) and Periodic Acid Schiff (PAS) were also done.

The ages of the children ranged from seven years to sixteen years. There were eight female children and four male children. All of them came from lower socioeconomic strata. They complained of foreign body sensation and pain prior to the appearance of conjunctival stone in the lower fornix. The stones ranged in size from 1mm to 1cm in different sizes, shapes and color varying from yellowish to white mottled and black to jet black. [Fig.1]



Fig-1: Gross appearance of stones formed in lower conjunctival fornix meas 1mm to 1 cm

General examination revealed moderately built and nourished children with no major illness. Examination of the eye was unremarkable except for mild to moderate inflammation of the lower conjunctival fornix. Visual acuity and fundus examination were normal. Routine investigations such as CBP, hematocrit, ESR and urine examination were within normal limits, but for mildly elevated eosinophil count in four children. Cytology smears of the conjunctival material showed cellular debris consisting of acute inflammatory cells and squamous epithelial cells. A refractile tangled mass of slender branched hyphae along with few conidia and areas of dark brown to black pigment deposits were seen. Clumps of needle shaped crystals and occasional dumbell shaped crystals were seen. Smears from the crushed stone material also showed crystalline material, fungal hyphae and pigment deposits [Fig.2] Diagnosis of *Aspergillus niger*was made on cytology. Microbiological culture of the stone on Sabouraud's Dextrose Agar showed fungal growth [Fig. 3].



Fig-2: Cytology: Refractile hyphae, pigmentdeposits and crystalline material (H & E) 1000x' is the completesentence



Fig-3: Fungal growth seen on culture

The fungus was identified as *A. niger* on microscopic examination with Lactophenol Cotton Blue (LPCB) stain [Fig.4]. No bacterial growth was seen. Biochemical analysis of the stones showed that they were composed of compounds of calcium. Tissue sections stained with hematoxylin and eosin revealed

moderate to severe non-specific inflammation of the conjunctival submucosal stroma, refractile branched hyphae, pigment deposits and crystals. Special stains, GMS and PAS revealed fungal hyphae with acute angle branching and conidia within the conjunctival epithelial layers and superficial stroma [Fig. 5].



Fig-4: HPE Conjunctival biopsy showing branched fungal hyphae & conidia (GMS 1000x)



Fig-5: LPCB stain 400x shows Aspergillus Niger

Table-1: The Microbiolo	ogy, Cytology, Histopathology & Biochemical findings are	e as shown

Diagnostic Test	Findings	Diagnosis
Microbiology Culture	Positive for fungal growth.	
Microscopy	Species identified on microscopy with LPCB stain	Aspergillus niger
	(Lactophenol cotton blue)	
Cytopathology	Tangled mass of refractile hyphae, Calcium oxalate	Aspergillus niger
(H & E)	crystals and dark brown pigment deposits	
Histopathology	Sub mucosal inflammation, Refractile branched fungal	Aspergillusniger
(H & E)	hyphae and conidia, pigment deposits and crystals	
Histopathology	Fungal hyphae with acute angle branching and conidia	Aspergillus niger
(Special stains -GMS & PAS)	within the epithelial layers & superficial stroma	
Biochemical analysis of stones	Calcium compounds	

### DISCUSSION

Aspergillosis occurs in immune compromised individuals though; cases of invasive aspergillosis occurring in immunocompetent individuals have been reported in literature. A case of cutaneous aspergillosis caused by *A. niger* has been reported in an immune competent patient [1].Another case of extrapulmonary disseminated aspergillosis and a case of maxillary sinus mycetoma both due to *A. niger*occurring in immunocompetent individuals are reported in literature [3, 4]. *A. niger* associated with otomycosis, cutaneous infections and pulmonary infections [5, 6].In the ocular tissues, aspergillus species have been implicated in a wide variety of infections characterized by either slow and asymptomatic infection or rapid uncontrollable progression [7].There are rare reports of *A. niger* infection complicating orbital exenteration in immunocompetent patients [8, 9].Patients in our study did not have any evidence of immunosuppression. *Aspergillusniger* prefers to grow in moist and warm environment and is commonly found in soil and plants [8,9].Hospital kitchens have been reported to be a source of this fungus [5]. In the present study, isolation of *A. niger* from the conjunctiva of children probably

suggests contamination due to poor personal hygiene and crowded environment.

A case of multiple white dacryoliths due to Aspergillusfumigatus is reported in literature [7]. There are isolated cases of conjunctival lithiasis in children reported in the news and media in India [10]. There are also isolated cases reported in the media from other countries such as Pakistan. Yemen and Nepal. However literature shows to the best of knowledge, only a single case reported by Lakshmi Narain in 1981[11]. Biochemical analysis in that case showed that the stones were composed of different compounds of calcium which is similar to the present study. Conjunctival biopsy in that case showed submucosal granulation tissue with perivascular calcification, whereas in the present study biopsy showed submucosal inflammation, branched fungal hyphae, black pigment deposits and calcium oxalate crystals. Deposition of calcium oxalate crystals in tissue is known to have strong association with aspergillus infections and more specific for A. niger strain [12]. The presence of calcium oxalate and black pigment deposits in lung parenchyma is considered as evidence of A. niger infection [13]. There was heavy calcium oxalate deposition on pathological examination in cases of A. niger causing invasive pulmonary aspergillosis [14, 15]. In the study of Choet al. the pathological specimen obtained by a transbronchial lung biopsy revealed numerous calcium oxalate crystals within the destructive lung lesions, but no fungal elements were identified [16]. The association of calcium oxalate crystals and Aspergillosis was first described in 1973 by Nime and Hutchins [12, 17]. Oxalic acid generates local oxidants that cause cell injury and tissue destruction, including blood vessel destruction [16, 17]. Although calcium oxalate is not always detected in patients with Aspergillus infection, its presence is considered as characteristic of A. niger infection [13, 17, 18]. The presence of calcium oxalate crystals should be considered as true sign of infection rather than colonization or contamination [12]. Identification of calcium oxalate crystals in sputum, fluids or tissue specimens should raise the possibility of aspergillosis caused by A. niger[19, 20]. Also the presence of black pigment with calcium oxalate crystals could be a pivotal clue to diagnosis even in the absence of fungal hyphae or conidia [13, 21]. Wehmer first described oxalic acid as a fermentation product of A. niger in 1981[13, 16, 17]. Both Aspergillusniger and fumigatus produce oxalic acid which precipitates as calcium oxalate by reacting with calcium ions in tissue fluids or blood [12, 13, 16, 20]. Oxalic acid is a mycotoxin released by the fungus, formed as a side product of TCA cycle by enzymatic hydrolysis of oxaloacetate by oxaloacetate acetylhydrolase [13,16].

Having established *A. niger* as the underlying cause of stone formation, the exact mechanism of conjunctival stone by *A.niger* formation remains to be determined. Studies on the pathogenesis of renal stone

formation are reviewed in order to understand the mechanism of stone formation. The pathogenisis of renal stone formation is a multistep process and includes nucleation, crystal growth, crystal aggregation and crystal retention [22].Nucleation is the formation of a solid crystal phase in a solution [23-25]. The process of nucleation in a pure solution is known as homogenous nucleation [23]. The process of nucleation that occurs over an existing surface such as epithelial cells, cell debris, RBCs etc. is known as heterogenous nucleation. Experimental studies demonstrated that injury from free radicals may result in sloughed membrane fragments providing a suitable nidus for nucleation. Stones result from a phase change in which dissolved salts condense into solids and this transformation is influenced by supersaturation. Crystal growth is the next step where several atoms or molecules in a supersaturated liquid start forming clusters. Crystal growth is determined by the molecular size and shape of crystals, physical properties of the material, supersaturation levels and pH. Epitaxy is a process where material of one crystal type is precipitated upon the surface of another whose lattice diemensions are almost identical [26]. Epitaxy is clinically important in stone formation. The next step is crystal aggregation in which crystals in solution stick together and form larger particle. Aggregation of particles in solution is determined by a balance of forces, some with disaggregating and some with aggregating effects. A small interparticle distance favors aggregation and the main force that inhibits aggregation is the repulsive electrostatic surface charge [22]. Crystal aggregation is more important than nucleation and growth because aggregation occurs within seconds [27]. Urolithiasis requires formation of crystals followed by their retention and accumulation in kidney by association of crystals to the renal epithelial cells [28]. Typically, supersaturation is not reached until the distal nephron, but in hyperoxaluric states, supersaturation for calcium oxalate is reached in the proximal tubule itself, which leads to crystal formation and contributes to stone formation [29].

Assuming similar mechanism for conjunctival stone formation, what causes instantaneous formation of conjunctival stones in comparison with the prolonged time taken for formation of renal stones is analysed. Urine contains several stone promoting factors and inhibiting factors, the imbalance between which has been suggested in the formation of renal stones. Calcium, sodium, oxalate, urate, cystine, low urinary pH and low urinary flow are some of the promoting factors for urinary stone formation. Cell debris, epithelial cell surfaces, protein aggregates and other crystals also act as promoters. Inhibitory factors include citrate, magnesium, pyrophosphate, Urinary protease prothrombin fragment 1, inhibitor, glycosaminoglycans, osteopontin, renal lithostathine and high urinary flow. These inhibitors cause growth inhibition of the stone at the level of nucleation,

adherence. Although aggregation or cell crystallogenesis is essential to stone formation, calcium oxalate stone growths in urine are sluggish [30]. To know the role of tear fluid in the formation of conjunctival stones the composition of tear fluid is studied. The major components of tear fluid are water, electrolytes, proteins (lysozyme, lactoferrin, lipocalin), secretory Ig A, albumin, IgG, lipids, Mucins, Defensins, collecctins et al. [31]. The electrolytes are principally Na<sup>+</sup>, K<sup>+</sup>, Cl<sup>-</sup>, and HCO<sup>-</sup> with lower levels of Mg<sup>2+</sup> and Ca<sup>2</sup>. The pH of the tear fluid in normal persons in the study of Norn MS was found to be 6.93+/- 0.24[32].

The following factors may possibly explain the rapidity with which stones form in the conjunctiva. Normally, tears constitute a thin film of fluid. Release of oxalic acid from Aspergillusniger leads to high oxalic acid concentration in the conjunctival sac which complexes with the calcium present in the small amount of tear fluid and achieves supersaturation levels quickly. This facilitates phase change and calcium oxalate crystal growth. Reduced tear secretion and changes in tear pH may contribute. The growth and aggregation of crystals into stones is potentiated by promoters of stone formation present in the tear fluid such as calcium, sodium, protein and surfaces of conjunctival epithelial cells. Presence of crystals and cell debris also act as promoters of stone formation. Compared to urine which has a large volume and numerous inhibitors of stone formation, tear fluid which is minute in volume seems to have very few inhibitors. Epitaxy seems to contribute to the formation of conjunctival stones. However further research to determine the molecular size and lattice diemensions of the crystals and factors determining crystal aaggregation need to be undertaken for clearer and more precise understanding of the mechanism of conjunctival stone formation.

# CONCLUSIONS

The cytohistological findings in correlation with mirobiological culture in this study revealed the presence of Aspergillusniger, both in the conjunctival tissues and in the material formed in the conjunctiva. The biochemical analysis of the stones revealed calcium compounds. Oxalic acid is a mycotoxin released by A. niger which complexes with calcium in tear fluid to form calcium oxalate crystals. Crystallogenesis is essential for stone formation. Factors such as minute tear fluid volume, the pH of tear fluid and calcium levels facilitate supersaturation leading to high oxalic acid concentration in the conjunctival sac contributing to rapid formation of conjunctival stones. Further research is needed for more precise understanding of the mechanism of conjunctival stone formation. This study helps to specifically treat this mysterious disorder which has remained an enigma for physicians since decades, with topical and oral antifungal agents. Voriconazole is the drug of choice for treatment of Aspergillusniger infections.

## REFERENCES

- Mohapatra S, Xess I, Swetha JV, Tanveer N, Asati D, Ramam M, Singh MK. Primary cutaneous aspergillosis due to Aspergillus niger in an immunocompetent patient. Indian journal of medical microbiology. 2009 Oct 1;27(4):367.
- Person AK, Chudgar SM, Norton BL, Tong BC, Stout JE.*Aspergillus niger:* an unusual cause of invasive pulmonary aspergillosis.J Med Microbiol. 2010 Jul;59(Pt 7):834-8.
- 3. Disseminated Aspergillosis due to *Aspergillus niger* in Immunocompetent Patient: A Case Report Case Rep Infect Dis
- 4. Zaman SU, Sarma DP. Maxillary sinus mycetoma due to Aspergillus niger. The Internet Journal of Otorhinolaryngology. 2007;6(1).
- Araiza J, Canseco P, Bonifaz A. Otomycosis: clinical and mycological study of 97 cases. Revue de laryngologie-otologie-rhinologie. 2006;127(4):251-4.
- Loudon KW, Coke AP, Burnie JP, Shaw AJ, Oppenheim BA, Morris CQ. Kitchens as a source of Aspergillus niger infection. Journal of hospital Infection. 1996 Mar 1;32(3):191-8.
- Comez AT, Komur B, Akcali A, Otkun MT. Ocular aspergillosis: Obtaining a specimen is crucial for diagnosis. A report of three cases. Medical mycology case reports. 2012 Jan 1;1(1):39-41.
- 8. So WL, Hardy TG. Aspergillus niger Infection of an Orbital Exenteration Socket Can Be Treated with Oral Itraconazole. Case reports in ophthalmological medicine. 2012;2012.
- Ugurlu S, Maden A, Sefi N, Sener G, Yulug N. Aspergillus niger infection of exenterated orbit. Ophthalmic Plastic & Reconstructive Surgery. 2001 Nov 1;17(6):452-3.
- 10. Available from https://www.youtube.com/watch?v=HI0Fp3VmP2 U
- 11. Narain L. Abnormal conjunctival lithiasis. Indian journal of ophthalmology. 1981 Jul 1;29(3):253.
- 12. Vakil RM, Patrawalla A, Cohen Z. Pulmonary Oxalosis as a Manifestation of Aspergillus niger Infection. Chest. 2010 Oct 1;138(4):110A.
- 13. Oda M, Saraya T, Wakayama M, Shibuya K, Ogawa Y, Inui T, Yokoyama E, Inoue M, Shimoyamada H, Fujiwara M, Ota T. Calcium oxalate crystal deposition in a patient with Aspergilloma due to Aspergillus niger. Journal of thoracic disease. 2013 Aug;5(4):E174.
- 14. Kimmerling EA, Fedrick JA, Tenholder MF. Invasive Aspergillus niger with fatal pulmonary oxalosis in chronic obstructive pulmonary disease. Chest. 1992 Mar 1;101(3):870-2.
- 15. Nakagawa Y, Shimazu K, Ebihara M, Nakagawa K. Aspergillus niger pneumonia with fatal pulmonary oxalosis. Journal of Infection and Chemotherapy. 1999 Jun 1;5(2):97-100.

- 16. Cho GJ, Ju JY, Park KH, Choi YD, Kim KS, Kim YI, Kim SO, Lim SC, Kim YC, Park KO, Nam JH. Pulmonary Oxalosis Caused by Aspergillus Niger Infection. Tuberculosis and Respiratory Diseases. 2003 Nov 1;55(5):516-21.
- 17. Nime FA, Hutchins GM. Oxalosis caused by aspergilus infection. Johns Hopkins Med J. 1973 Oct;133(4):183-94.
- Kurrein F, Green GH, Rowles SL. Localized deposition of calcium oxalate around a pulmonary Aspergillus niger fungus ball. American journal of clinical pathology. 1975 Oct 1;64(4):556-63.
- Mackowiak PA, Harigopal P, Bejarano P, Burke G, Dowdy LM. Birefringent crystals in a lung cavity.Clinical infectious diseases. 2005 Jun 15;40(12):1849-50.
- Procop GW, Johnston WW. Diagnostic value of conidia associated with pulmonary oxalosis: evidence of an Aspergillus niger infection. Diagnostic cytopathology. 1997 Oct;17(4):292-4.
- 21. Vaideeswar P, Sakhdeo UM. Pulmonary aspergilloma with renal oxalosis: fatal effect at a distance. Mycoses. 2009 May;52(3):272-5.
- 22. Basavaraj DR, Biyani CS, Browning AJ, Cartledge JJ. The role of urinary kidney stone inhibitors and promoters in the pathogenesis of calcium containing renal stones. EAU-EBU update series. 2007 Jun 1;5(3):126-36.
- 23. Finlayson B. Physicochemical aspects of urolithiasis. Kidney Int 1978;13:344–60.

- 24. Kok DJ. Intratubular crystallization events. World J Urol 1997;15:219–28.
- 25. Khan SR, Byer KJ, Thamilselvan S, Hackett RL, Mccormack WT, Benson NA, Vaughn KL, Erdos GW.Crystal-cell interaction and apoptosis in oxalate-associated injury of renal epithelial cells. Journal of the American Society of Nephrology: JASN. 1999 Nov;10:S457-63.
- 26. Lonsdale K. Epitaxy as a growth factor in urinary calculi and gallstones. Nature 1968;217:56–8.
- Hess B, Zipperle L, Jaeger P. Citrate and calcium effects on Tamm-Horsfall glycoprotein as a modifier of calcium oxalate crystal aggregation. Am J Physiol 1993;265:F784-91.
- Asselman M, Verkoelen CF. Crystal-cell interaction in the pathogenesis of kidney stone disease. Current opinion in urology. 2002 Jul 1;12(4):271-6.
- 29. Lorenz EC, Michet CJ, Milliner DS, Lieske JC. Update on oxalate crystal disease. Current rheumatology reports. 2013 Jul 1;15(7):340.
- Finlayson B, Reid F. The expectation of free and fixed particles in urinary stone disease.Investigative urology. 1978 May;15(6):442-8.
- Tiffany JM. Tears in health and disease.Eye. 2003 Nov;17(8):923.
- 32. Norn MS. Tear fluid pH in normals, contact lens wearers, and pathological cases.Acta ophthalmologica. 1988 Oct;66(5):485-9.